

Review Article

Role of Redox Signaling in Neuroinflammation and Neurodegenerative Diseases

Hsi-Lung Hsieh¹ and Chuen-Mao Yang²

¹ Department of Nursing, Division of Basic Medical Sciences, Chang Gung University of Science and Technology, Taoyuan, Taiwan

² Department of Physiology and Pharmacology and Health Aging Research Center, College of Medicine, Chang Gung University, 259 Wen-Hwa 1st Road, Kwei-San, Taoyuan, Taiwan

Correspondence should be addressed to Chuen-Mao Yang; chuenmao@mail.cgu.edu.tw

Received 11 September 2013; Revised 30 October 2013; Accepted 21 November 2013

Academic Editor: Sulagna Das

Copyright © 2013 H.-L. Hsieh and C.-M. Yang. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Reactive oxygen species (ROS), a redox signal, are produced by various enzymatic reactions and chemical processes, which are essential for many physiological functions and act as second messengers. However, accumulating evidence has implicated the pathogenesis of several human diseases including neurodegenerative disorders related to increased oxidative stress. Under pathological conditions, increasing ROS production can regulate the expression of diverse inflammatory mediators during brain injury. Elevated levels of several proinflammatory factors including cytokines, peptides, pathogenic structures, and peroxidants in the central nervous system (CNS) have been detected in patients with neurodegenerative diseases such as Alzheimer's disease (AD). These proinflammatory factors act as potent stimuli in brain inflammation through upregulation of diverse inflammatory genes, including matrix metalloproteinases (MMPs), cytosolic phospholipase A₂ (cPLA₂), cyclooxygenase-2 (COX-2), and adhesion molecules. To date, the intracellular signaling mechanisms underlying the expression of target proteins regulated by these factors are elusive. In this review, we discuss the mechanisms underlying the intracellular signaling pathways, especially ROS, involved in the expression of several inflammatory proteins induced by proinflammatory factors in brain resident cells. Understanding redox signaling transduction mechanisms involved in the expression of target proteins and genes may provide useful therapeutic strategies for brain injury, inflammation, and neurodegenerative diseases.

1. Introduction

In general, inflammation is a protective response to various cell and tissue injuries. The purpose of this process is to destroy and remove the detrimental agents and injured tissues, thereby benefiting tissue repair. When this helpful response is uncontrolled, the effect initiates excessive cell and tissue damages that result in destruction of normal tissue and chronic inflammation [1–3]. Moreover, the brain inflammatory diseases, including Alzheimer's disease (AD) and Parkinson's disease (PD), are characterized by “redox state” imbalance and chronic inflammation, a major cause of cell damage and death. Reactive oxygen species (ROS) are widely recognized as key mediators of cell survival, proliferation, differentiation, and apoptosis [4, 5]. Excessive

production of ROS (termed “oxidative stress”) by mitochondria and NADPH oxidase (Nox) is usually thought to be responsible for tissue injury associated with a range of brain injury, inflammation, and degenerative diseases such as AD [5–8]. Moreover, many of the well-known inflammatory target proteins, including matrix metalloproteinase-9 (MMP-9), cytosolic phospholipase A₂ (cPLA₂), cyclooxygenase-2 (COX-2), inducible nitric oxide synthase (iNOS), and adhesion molecules, are associated with oxidative stress (ROS generation) induced by proinflammatory factors such as cytokines, peptides, infections, and peroxidants [3, 5, 9]. Brain cells, especially neuroglial cells, are susceptible to the injurious effects of oxidative stress. Several studies have shown that brain cells like microglia and astrocytes induce and release diverse inflammatory mediators in response to

oxidative stress [9–11]. In addition, ROS act as a critical signaling molecule to trigger inflammatory responses in central nervous systems (CNS) through the activation of the redox-sensitive transcription factors, including nuclear factor- κ B (NF- κ B) and activator protein-1 (AP-1) [5, 9]. Thus, this review will focus on many general aspects of oxidative stress regulation and summarize the current progresses regarding the occurrence and effects of redox signals on CNS and their involvement in the expression of inflammatory target proteins in response to proinflammatory factors during brain inflammation. Moreover, the pharmacological interventions which protect against oxidative stress-induced neuroinflammation and neurodegenerative diseases will be discussed.

2. Role of Neuroglial Cells in CNS Physiological and Pathological Events

CNS consists of neurons and glial cells. Among glial cells, astrocytes constitute nearly 40% of the total CNS cell population in the adult human brain, and they maintain homeostasis in normal CNS. Astrocytes have also been proposed to exert a wide range of functions including guidance of the development and migration of neurons during brain development, production of growth factors, maintenance of the integrity of the blood-brain barrier (BBB), and participating in the immune and repairing responses to disease and brain injury [12, 13]. Microglial cells represent resident brain macrophages and can be transformed into activated immunocompetent antigen-presenting cells during the pathological process. An increased number of activated microglial cells have consistently been reported in PD, which may have a deleterious effect on dopaminergic neurons [14]. Astrocytes, as well as microglia, display an array of receptors involved in innate immunity, including Toll-like receptors (TLRs), nucleotide-binding oligomerization domains, double-stranded RNA dependent protein kinase, mannose receptor, and components of the complement system [10]. One common feature of a variety of neurodegenerative disorders is the presence of a large number of activated glial cells including astrocytes and microglia that involve the changes of morphology and expression of many inflammation-related proteins. Gliosis, especially astrogliosis, is characterized by astrocytic proliferation, extensive hypertrophy of the cell body, and functional changes, when stimulated with various factors including lipopolysaccharide (LPS), interleukin-1 β (IL-1 β), and tumor necrosis factor-(TNF- α) [15, 16].

Moreover, the cell-cell interactions between glial cells and neurons may be important in the regulation of brain inflammation and neurodegeneration. Many recent reports implicate that inflammation contributes to a wide variety of brain pathologies, apparently killing neurons via glia [10, 11, 17]. Thus, the activated glial cells, especially microglia and astrocytes, are thought to play a critical role in the pathogenesis and progression of neurodegeneration (Figure 1). Previously, many reports have shown that microglial cells may be a major inflammatory cell of the brain [14]. The activated microglia produce several inflammatory mediators including COX-2/prostaglandins (PGs), iNOS/nitric oxide (NO), or

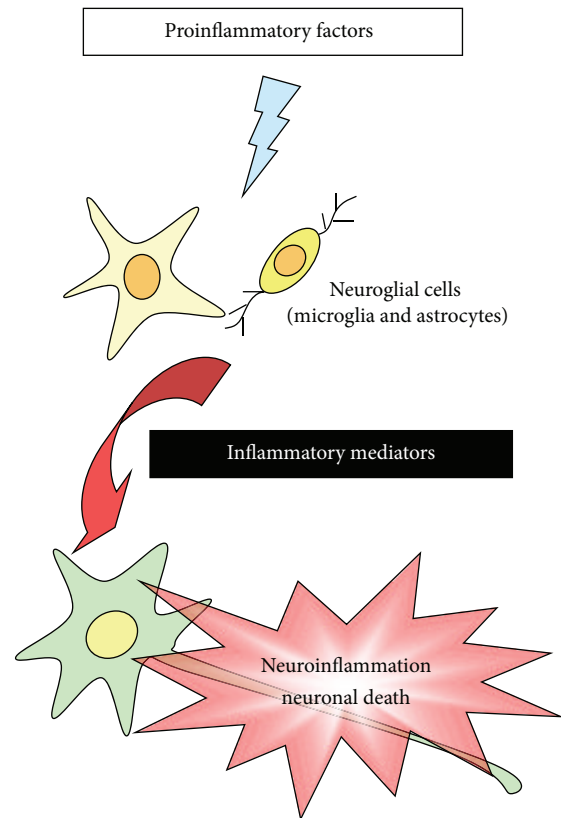


FIGURE 1: Schematic presentation of the interaction of the brain cells, including neurons and glial cells. In the central nervous system (CNS), proinflammatory factors induce the expression of various inflammatory mediators in neuroglial cells, particularly microglia and astrocytes. These induced inflammatory mediators from glial cells may cause the neuroinflammation or neuronal death, and then leading to neurodegenerative disorders.

cytokines as well as neurotoxic substances, which are thought to be responsible for brain injuries and diseases including trauma, AD, and neural death due to the exposure of LPS, interferon- γ , or β -amyloid [18, 19]. Although most studies have demonstrated that microglial cells play an important role in neuroinflammation and neurodegeneration, accumulating evidence has also demonstrated the characteristic changes of astrocytes in neurodegenerative diseases such as dementia [10, 11, 20]. Recently, we have demonstrated the upregulation of several inflammatory mediators including MMP-9, cPLA₂, COX-2, iNOS, and oxidative stress by various proinflammatory factors such as cytokines (e.g., IL-1 β), peptides (e.g., bradykinin (BK) or endothelin-1 (ET-1)), infections (e.g., bacteria or virus), and peroxidants (e.g., oxidized low-density lipoprotein (oxLDL)) in rat brain astrocytes [21–29]. More recent data indicated that multiple factors including ROS, MMP-9, and heme oxygenase-1 (HO-1)/carbon monoxide (CO) from BK-challenged brain astrocytes may contribute to the neuronal cell apoptosis [30]. Together these results implicate that activated neuroglial cells, especially astrocytes, play a key role in the pathogenesis of the CNS inflammation leading to neurodegenerative diseases (Figure 1).

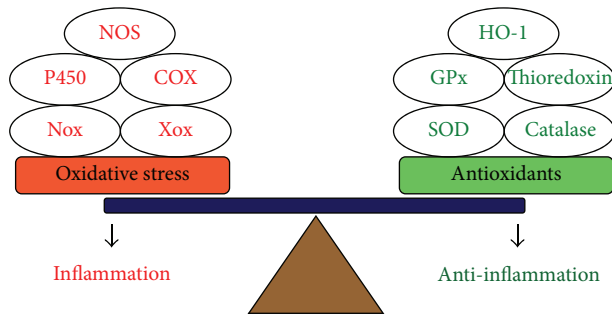


FIGURE 2: Oxidative stress and antioxidants imbalance in inflammation. In inflammation, the balance appears to be tipped in favor of increased oxidative stress by various specialized enzymes, including Nox, Xox, P450, COX, or NOS, either because of excessive ROS release or inflammatory mediators leading to the amplification of the proinflammatory effects. In contrast, induction of several antioxidants, such as SOD, catalase, GPx, thioredoxin, or HO-1, may reduce ROS generation and attenuate the inflammatory response (anti-inflammation). Nox: NADPH oxidase; Xox: Xanthine oxidase; P450: P450 enzyme; COX: cyclooxygenase; NOS: nitric oxide synthase; SOD: superoxide dismutase; GPx: glutathione peroxidase; HO-1: heme oxygenase-1.

3. Role of Oxidative Stress (Redox Signaling) in the Brain Inflammation and Neurodegenerative Diseases

In CNS inflammation, various proinflammatory factors may cause the development of an oxidative stress and antioxidants imbalance, which induces redox signal-dependent expression of genes for inflammatory mediators or protective antioxidants (Figure 2). The oxidative stress (i.e., ROS and reactive nitrogen species (RNS)) is produced by various enzymatic reactions and chemical processes or directly inhaled. ROS that are particularly responsible in oxidative stress include superoxide anion ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2), and hydroxyl radical ($\cdot OH$). Furthermore, the RNS include nitric oxide (NO) and peroxynitrite ($ONOO^-$). These oxidative stresses (i.e., ROS/RNS) are essential for many physiological functions at low concentrations [2–6] and killing invading microorganisms [31]. However, several lines of evidence have suggested that the pathogenesis of human diseases is attributed to increased oxidative stress [2, 31]. Moreover, oxidative stress has been shown to mediate the pathogenesis of neurodegenerative diseases, including PD [6], AD [32], and cerebrovascular disorders such as stroke [31]. There are several major sources of ROS/RNS generation in the cells, including Nox, Xanthine oxidase (Xox), P450 enzymes, COX, and NOS (Figure 2), which contribute to several physiological and pathological functions including brain inflammation and neurodegeneration [8]. The physiological role of ROS/RNS (along with $O_2^{\cdot-}$ and NO) also extends to the control of vascular tone in the brain, which is tightly modulated by the metabolic activity within neurons [6, 33]. Particularly in the brain, even small redox imbalances can

be deleterious. Recently, accumulating evidence attributes the cellular damage in the CNS degenerative disorders to oxidative stress [5–9], suggesting that oxidative stress is an early event in AD [32]. Oxidative stress may be responsible for brain inflammatory disorders, which cause deleterious effects during CNS pathogenesis [34]. Furthermore, several reports have shown that ROS levels are increased with age in several major organs including brain [32]. Abnormally elevated ROS is implicated in age-related long-term potentiation (LTP) impairment [35]. ROS further induce expression and activation of proinflammatory factors or inflammatory mediators during brain injury and inflammation. Under various pathological conditions, excessive amounts of ROS can damage DNA, lipids, proteins, and carbohydrates leading to impairing cellular functions and enhancing inflammatory reactions [34, 36]. In brains of AD patients, cellular and animal models of AD, the elevated levels of these oxidative stress-modified molecules are also detected [32]. Recently, increasing evidence attributes the cellular damage in neurodegenerative disorders such as AD and PD to oxidative stress that leads to generation of ROS associated with brain inflammatory disorders [2, 6]. Thus, these results indicate that oxidative stress (i.e., ROS production) plays an important role in CNS inflammation and neurodegenerative disorders (Figure 4).

Oxidative stress activates several intracellular signaling cascades that may have a deleterious effect on the cellular homeostasis. The molecular mechanisms associated with ROS production (e.g., mitochondrial dysfunction and Nox activation) and its influences have been investigated in various models of chronic inflammation and neurodegenerative disorders [9]. Recently, there are extensive pieces of literature supporting a role of mitochondrial dysfunction and oxidative damage in the pathogenesis of AD [5, 37], and ROS are associated with neuroinflammatory and neurodegenerative processes [9, 17, 32]. Several proinflammatory factors (e.g., LPS and BK) have been shown to induce the expression and activation of various inflammatory mediators via a ROS-dependent manner in brain cells [25, 36]. In microglial cells, ROS, as a major signaling molecule, mediate microglial activation induced by proinflammatory mediators such as $A\beta$ or LPS [38, 39]. However, the roles of oxidative stress that contribute to these events are not well characterized in brain cells including astrocytes. Our recent reports have demonstrated that ROS signals contribute to the expression of many inflammatory genes (e.g., MMP-9) by several proinflammatory factors, including BK [25], LTA [27], and TGF- β 1 [40] in brain astrocytes. More recent result indicates that ROS generation from BK-challenged astrocytes contributes to neuronal apoptosis through a caspase-3-dependent manner [30]. Although oxidative stress is implicated as a causative factor in neurodegenerative disorders, the signaling pathways linking ROS production with neuronal cell death are not well characterized [6]. Hence, there are several targets and signals that need to be identified and explored for the development of therapeutic strategies in the future.

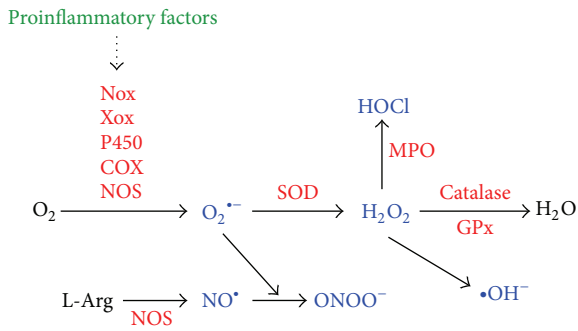


FIGURE 3: Major pathways of reactive oxygen (nitrogen) species generation and metabolism. Several proinflammatory factors can stimulate $O_2^{\bullet-}$ generation through activation of several specialized enzymes, such as the Nox, Xox, P450, COX, or NOS. SOD then converts the $O_2^{\bullet-}$ to H_2O_2 , which is then converted into the highly reactive $\bullet OH$ or has to be rapidly removed from the system that is generally achieved by catalase or peroxidases, such as the GPx. Further, $O_2^{\bullet-}$ can be either converted into $ROO\bullet$ or can react with NO to yield $ONOO^-$. NO is mostly generated by L-Arg via NOS. H_2O_2 can be converted to HOCl by the action of MPO, myeloperoxidase. O_2 : molecular oxygen; H_2O : water; $O_2^{\bullet-}$: superoxide radical anion; $\bullet OH$: hydroxyl radical; $ROO\bullet$: peroxy radical; H_2O_2 : hydrogen peroxide; $ONOO^-$: peroxynitrite; NO: nitric oxide; L-Arg: L-arginine; HOCl: hypochlorous acid.

4. Redox Signaling and Proinflammatory Factors in Brain Inflammation and Neurodegenerative Diseases

The senile and neuritic plaque of AD are accompanied by inflammatory responses in activated glial cells (i.e., astrocytes and microglia). In CNS, several cytokines and inflammatory mediators produced by activated glia have the potential to initiate or exacerbate the progression of neuropathology [41]. Moreover, traumatic injury to CNS results in the production of inflammatory cytokines via intrinsic (brain cells) and extrinsic means (by infiltrating macrophages and other leukocytes). The expression of many inflammatory mediators including cytokines, MMPs, cPLA₂, COX-2, and iNOS has been shown to be regulated by various extracellular stimuli such as proinflammatory cytokines (e.g., IL-1 β and TNF- α), peptides (e.g., BK, ET-1, and A β), infections (e.g., bacteria and virus), peroxidants (e.g., oxLDL and H₂O₂), and other stresses (e.g., TGF- β) in neuronal and neuroglial cells [4–9, 42] (Figure 4).

4.1. Cytokines. IL-1 β and TNF- α are two of the inflammatory cytokines significantly elevated in neurodegenerative diseases such as AD, and they play a central role in initiating and regulating the cytokine cascades during inflammatory responses [43]. IL-1 β is a pleiotropic cytokine and classified as a dominant injury biomarker. Furthermore, several studies have shown that the level of IL-1 β is elevated in the cerebrospinal fluid (CSF) of patients with AD, traumatic brain injury [44], and stroke [45]. Thus, IL-1 β plays an important role in both acute and chronic neurodegenerative diseases.

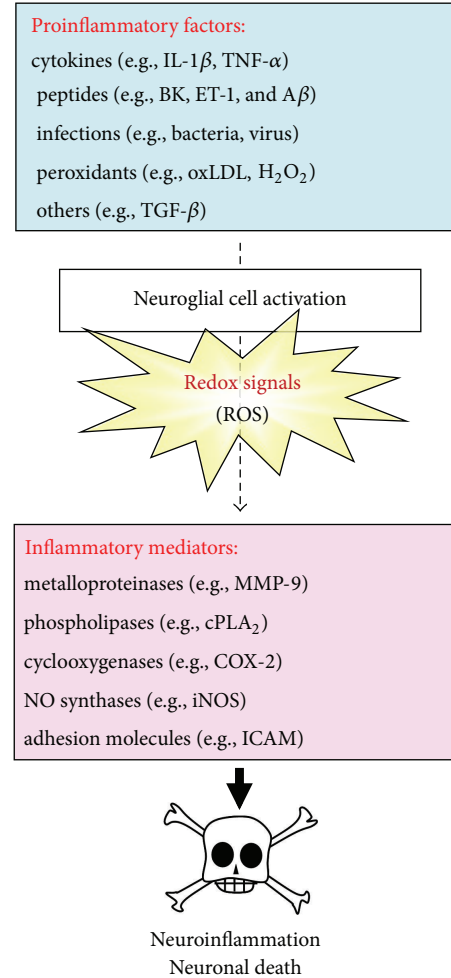


FIGURE 4: Schematic representation of the redox signals (ROS production) and their role in the development of neuroinflammation and neuronal death. Many of the well-known inflammatory target proteins, such as MMP-9, ICAM-1, VCAM-1, COX-2, and cPLA₂, can be upregulated by various proinflammatory factors, including cytokines, peptides, bacterial or viral infection, peroxidants, via a ROS signal-dependent manner in neuroglial cells. These inflammatory mediators can cause neuroinflammation and neuronal death. IL-1 β : interleukin-1 β ; TNF- α : tumor necrosis factor- α ; BK: bradykinin; ET-1: endothelin-1; A β : β -amyloid; oxLDL: oxidized low-density lipoprotein; H₂O₂: hydrogen peroxide; TGF- β : transforming growth factor- β ; MMP-9: matrix metalloproteinase-9; cPLA₂: cytosolic phospholipase A₂; COX-2: cyclooxygenase-2; iNOS: inducible nitric oxide synthase; ICAM: intercellular adhesion.

The effects of IL-1 β on ROS generation have been reported to be associated with brain inflammatory disorders, cancers, and myocardial remodeling [46, 47]. ROS generation by IL-1 β leads to the expression of several inflammatory genes like MMP-9 which may increase BBB permeability, recruit immune cells infiltrating through BBB into the tissues, and subsequently result in brain inflammation and edema during brain injury [6, 34]. ROS may also act as an inflammatory signaling factor mediated microglial activation induced by IL-1 β [39]. Moreover, in culture of glia/neuron, IL-1 β induces

neurotoxicity through the release of free radicals [48]. In addition, TNF- α is also produced in response to oxidative stress and A β . In brain, TNF- α is produced by microglia and its overproduction has been linked with neuronal cell death [49]. These studies indicate that cytokines, especially IL-1 β and TNF- α , contribute to the CNS inflammation and neurodegenerative diseases through redox signalings.

4.2. Peptides. AD is defined by progressive impairments in memory and cognition and by the presence of extracellular neuritic plaques (A β) and intracellular neurofibrillary tangles (tau protein) [5, 32]. Among these molecules, A β is an insoluble fibrous protein and aggregates sharing specific structural traits. It arises from at least 18 inappropriately folded versions of proteins and polypeptides present naturally in the body. The misfolded structures alter their proper configuration such that they erroneously interact with other cell components forming insoluble fibrils. A β has been associated with the pathology of more than 20 human diseases including AD. Abnormal accumulation of amyloid fibrils in brain may play a role in neurodegenerative disorders. Although A β peptide is neurotoxic species implicated in the pathogenesis of AD, mechanisms through which intracellular A β impairs cellular properties and produces neuronal dysfunction remain unclear. Accumulating evidence has indicated that A β can stimulate the production of free radicals [50]. Interestingly, intracellular A β is present in mitochondria from brains of transgenic mice with targeted neuronal overexpression of mutant human amyloid precursor protein and AD patients. Importantly, mitochondria-associated A β , principally A β ₁₋₄₂, was detected as early as 4 months, before extensive extracellular A β deposits [51]. Moreover, activation of Nox by A β ₁₋₄₂ results in ROS production in rat primary culture of microglial cells [52]. In mouse models of plaque formation, oxidative stress occurs prior to A β deposition in a Tg2576 APP transgenic mice [53]. Moreover, increased levels of oxidative damage occur in individuals with mild cognitive impairment (MCI), which is often believed to be one of the earliest stages of AD [54]. Additionally, glial HO-1 expression in the MCI temporal cortex and hippocampus is also significantly greater than that of the nondemented group [55]. These results support A β -induced redox signaling serving as an early event that leads to the development of the CNS pathological features such as AD. Moreover, glial cells may play a key role in the events.

In addition to A β peptide, BK and related peptides are produced and released during trauma, stroke, and neurogenic inflammation [56]. All these pathological processes may be involved in tissue remodeling, which were regulated by MMPs. Moreover, astrocytes possess receptors for numerous transmitters such as glutamate and BK [57]. These peptides mediate several inflammatory responses including increasing vasodilatation and vascular permeability, promotion of fluid secretion and ion transport, and eliciting itching and pain at the sites exposed to noxious stimuli. Thus, the elevated level of BK plays a key role in the initiation of inflammatory responses in target tissues, including CNS. It is well established that BK interacts with two BK receptor subtypes, including BK B1 and B2 [58]. Astrocytes are known to express

B2-type BK receptors and this type of receptors is found only on astrocytes type 1 [57]. The B2 BK receptor is a heterotrimeric G-protein-coupled receptor (GPCR) that can be coupled to intracellular signaling molecules via interaction with G_q protein [59]. Activation of BK receptors stimulates intracellular signaling molecules, including Ca²⁺, PKCs, and MAPKs, in several cell types including astrocytes [57-59]. Activation of these signaling pathways may lead to cell survival, proliferation, differentiation, and the expression of several inflammatory genes such as iNOS and MMP-9 [36, 60]. During brain injury, BK has been shown to induce the expression of several inflammatory genes by increasing ROS production [6, 34]. Moreover, Nox is expressed in astrocytes and contributes to ROS generation [61, 62]. In brain astrocytes, BK induces the expression of several inflammatory genes like MMP-9 by ROS-dependent signaling pathways [25]. Moreover, ROS released from BK-challenged brain astrocytes cause neuronal cell apoptosis [30]. These pieces of literature suggest that BK plays an important role in brain inflammation and neurodegenerative disorders.

Endothelial cells are known to produce vasotone mediators such as endothelins (ETs) and NO to maintain hemodynamic responses. The ETs are 21-amino acid vasoconstricting peptides produced primarily in the endothelium, which play a key role in vascular homeostasis and have been implicated in brain inflammatory diseases. Among the ET family, the bioactivity of ET-1 is mediated through potent vasoconstrictor and proinflammatory action in vascular diseases, including the heart, circulation system, and brain [63-66]. Two types of ET receptors, ET type A (ET_A) and type B (ET_B), are responsible for ET-1-triggered biological effects, which are mediated via G-protein-dependent processes [63-65]. In CNS, ET-1 also plays a substantial role in the normal development and CNS diseases. Both endothelial cells and astrocytes are potential sources of ET-1 release in response to hypoxic/ischemic injury of the brain [66]. On astrocytes, the ET_B receptors are predominantly expressed and modulate postinjury responses of astrocytes in CNS [67]. Circumstantial evidence has further demonstrated that overexpression of ET-1 has deleterious effects on astrocytes in ischemic brain [68]. Similarly, ET-1 causes hypertrophy of ET_B/GFAP-immunoreactive astrocytes, a typical characteristic of astrogliosis, in the normal optic nerve, leading to glial scar formation following CNS injury [68]. Endothelial ET-1 induces cytokine production such as IL-1 β released by astrocytes, which directly contributes to BBB breakdown during CNS inflammation [69]. These findings further imply the involvement of ET-1 in the CNS inflammation and diseases.

4.3. Infections. Bacterial infections have been shown to be involved in brain inflammation [70]. A well-known endotoxin from Gram-negative bacteria, LPS, regulates the expression of inflammatory proteins associated with inflammatory diseases. Many studies have also shown that ROS are the major signaling molecule which mediates microglial activation induced by inflammatory mediators, including LPS [71]. However, the signaling mechanisms of which activated brain

cells in response to Gram-positive bacterial infection remain undefined. Gram-positive bacterial infections of CNS occur in bacterial meningitis and brain abscess, being localized to the membranes surrounding the brain and in its parenchyma [72]. Lipoteichoic acid (LTA), an amphiphilic polymer, is embedded in-cell wall of Gram-positive bacteria [73]. The Gram-positive bacterium *Streptococcus pneumoniae* is the most common cause of acute bacterial meningitis worldwide [74], revealing a close relationship between LTA challenges and CNS diseases. For the initiation of LTA signaling, TLRs are believed to be responsible for LTA recognition challenged by Gram-positive bacteria such as *Staphylococcus aureus* and *Streptococcus pneumoniae* [75]. Upon binding to TLR heterodimers (i.e., TLR2/TLR1 or TLR2/TLR6 complex), LTA exerts a sequential activation of members of IL-1 receptor-associated kinase (IRAK) family and tumor necrosis factor receptor-associated factor 6 (TRAF6), mediated by a TLR adaptor protein MyD88. Ultimately, TLR signalings activate MAPK family and NF- κ B, leading to modulation of gene expression of cytokines and other inflammatory proteins [76]. Among the diverse cell types in CNS, glial cells such as astrocytes and microglia are regarded as targets in Gram-positive bacterial infection [77–79]. Several lines of evidence suggest a causal relationship between LTA challenges and the CNS diseases, which involves glial activation and TLR2 signalings [77–79]. TLR signalings in astrocytes have been shown to be involved in inflammatory responses in CNS [80], accompanied with upregulation of genes with inflammatory and proapoptotic effects [81]. The pathogenic progression involves glial activation and TLR2 signalings stimulated by LTA, which are linked to inflammatory neurodegeneration [82]. Additionally, LTA exhibits detrimental effects on brain cellular functions, including induction of apoptosis, production of oxidative stresses, and disruption of BBB following group B *Streptococcus* or *Staphylococcus aureus* challenge in CNS [82]. Although the effects of LTA on ROS generation have been reported in several cell types such as renal diseases [83], LTA-induced brain cell responses through the ROS signals are not well characterized. Recent report indicates that LTA-induced MMP-9 expression is mediated through Nox2-derived ROS generation in brain astrocytes [27]. These data suggest that targeting LTA and its specific signaling components could yield useful therapeutic targets for CNS inflammatory diseases upon infection with Gram-positive bacteria.

Moreover, increasing evidence has shown that viral infections such as Japanese encephalitis virus (JEV) and Enterovirus 71 (EV71) may contribute to several inflammatory responses in CNS [28]. Neurotropic viruses can cause massive neuronal dysfunction and destruction that lead to neurological diseases. EV71, a single-positive-stranded RNA virus, belongs to the Enterovirus B genus of the Picornaviridae family [84]. EV71 and Coxsackievirus A16 (CVA16) are the major causative agents of hand-foot-and-mouth disease (HFMD) that is usually mild exanthematous infection and self-limiting in the young children. However, EV71, but not CVA16, can progress to severe neurological diseases including fatal encephalitis, aseptic meningitis, and fatal neurogenic pulmonary edema [85]. Children under 5

years old of age group are susceptible to these infections and may develop permanent neurological sequelae or even succumb to such disorders [86]. In 1998, an EV71 outbreak infected more than 130,000 children resulted in 78 fatalities. Since then, EV71 infection has recurred every year in Taiwan and EV71 outbreaks have been periodically reported throughout the world, representing a major public health concern particularly in the Asia-Pacific regions including Taiwan, Malaysia, Singapore, Japan, and China [85, 87]. The emerging evidence suggests that ROS affect the interaction between host and viral pathogens. Recently, EV71 has been shown to induce oxidative stress-dependent viral replication in human neuroblastoma SK-N-SH cell line [88]. Similarly, JEV is a single-stranded, positive-sense RNA virus belonging to the family Flaviviridae. JEV is transmitted between animals and humans by culex mosquitoes [89]. After the bite of an infected mosquito, JEV amplifies peripherally producing transient viremia before entering into CNS [89]. The principal target cells for JEV are localized in CNS, including neurons and astrocytes [90]. Several lines of evidence suggest that JEV frequently causes severe encephalitis in the world, especially in Eastern and Southeastern Asia. The infection with JEV is characterized by clinical manifesting with fever, headache, vomiting, signs of meningeal irritation, and altered consciousness leading to high mortality [89, 90]. The generation of ROS plays an important role in diverse cellular functions including signal transduction, oxygen sensing and host defense during infection by viruses such as JEV [91]. In CNS, JEV infection has been shown to upregulate MMP-9 gene expression through ROS-dependent pathways in brain astrocytes [28]. These findings concerning JEV-induced expression of inflammatory genes in brain astrocytes imply that JEV might play a critical role in the brain inflammation and neurodegenerative diseases.

4.4. Peroxidants. Oxidative stress may cause production of several peroxidants such as oxidized lipoprotein. Clinical reports reveal that the patients with AD exhibit an increased oxidation of lipoproteins potentially toxic to neurons in CNS [92]. Among these, the oxidized low-density lipoprotein (oxLDL) is a well-known predominantly risk factor of atherosclerosis, which has been reported to participate in the progression of the CNS diseases. In CNS, oxLDL exhibits detrimental effects on brain cell functions, including induction of apoptosis, disruption of capillary homeostasis, and alteration of inflammatory protein activity in various brain cells [93]. Furthermore, in patients with cerebral infarction, oxLDL is present in brain parenchyma and stimulates astrocytes to secrete interleukin-6 [94] and may serve as an indicator to reflect the level of oxidative stress [95]. In brain astrocytes, oxLDL can induce MMP-9 expression and cell migration, which plays a critical role in the progression of inflammatory diseases and remodeling processes in target tissues, including CNS [29, 96]. These findings suggest that peroxidants like oxLDL might play a key role in the progression of the CNS diseases and also that targeting these peroxidants-stimulated signaling components may provide useful therapeutic strategies for brain inflammation and neurodegenerative diseases.

4.5. Others. In addition to these well-known factors, there are many factors that may also contribute to neuroinflammatory responses. Among these, TGF- β has been implicated to participate in the responses. TGF- β binds to two serine/threonine kinase receptors which consist of TGF- β RI and TGF- β RII. During ligand binding, TGF- β RII phosphorylates TGF- β RI and activates Smad-dependent intracellular signaling pathways and thus leads to expression of several genes [97, 98]. In addition to activation of Smad-dependent pathways, TGF- β can affect several signal transduction pathways in a Smad-independent manner, such as MAPKs [97, 98]. In human gingival and skin fibroblasts, both p38 MAPK and Smad3 cooperate in regulating TGF- β -induced MMP-13 expression, whereas ERK1/2 cooperates with Smad3 in regulating connective tissue growth factor expression [99]. Recently, increasing evidence has attributed the cellular damage in neurodegenerative disorders to oxidative stress leading to generation of ROS that are responsible for brain inflammation and neurodegenerative disorders [6, 34]. TGF- β can stimulate ROS production, which participates in the expression of diverse inflammatory genes such as MMPs in the processes of several human inflammatory diseases [100]. In brain astrocytes, TGF- β 1 has been shown to induce inflammatory protein expression via a ROS-dependent manner [40]. These results suggest that TGF- β 1 may play a key role in the process of brain inflammation and neurodegenerative diseases.

5. Role of Redox Signaling in the Regulation of Inflammatory Mediators

Neuroinflammation is an active defensive process against diverse insults, metabolic and traumatic injuries, infection, and neurodegenerative diseases. Although neuroinflammation serves as a neuroprotective mechanism associated with repair and recovery, it can also cause brain damage [101]. However, if inflammation in the brain is chronic or inappropriately controlled, it may become detrimental to neurons, thus representing one of the various pathological insults induced by various proinflammatory factors and by inflammatory mediators in CNS [101]. Experimental and clinical studies have shown that various inflammatory mediators are present in brain, CSF, and blood in brain injury. In particular, the histological analysis of human brain from individuals with brain disorder such as AD or epilepsy of various etiologies strongly suggests the existence of a chronic inflammatory state in the brain almost invariably associated with neuronal loss or reactive gliosis [102]. In experimental models of rodent brain seizures, a variety of inflammatory mediator mRNAs and protein levels are rapidly increased after the induction of seizures, including MMPs (e.g., MMP-9, especially), multiple forms of PLA₂ (e.g., cPLA₂), COX-2, NOS (e.g., iNOS), and adhesion molecules (e.g., ICAM-1 and VCAM-1) [102, 103]. After expression of these inflammatory mediators, several CNS damaging factors will be produced such as cytokines shedding by MMPs, arachidonic acid (AA)/PGE₂ releasing by cPLA₂/COX-2 system, and NO generation by NOS [102, 103]. Herein, we reviewed the role

and mechanism of these inflammatory mediators in the brain inflammation and neurodegeneration and whether oxidative stress plays a crucial role in these events.

5.1. Matrix Metalloproteinases. MMPs are a large family of zinc-dependent endopeptidases, which play an important role in the turnover of extracellular matrix (ECM) and pathophysiological processes [104]. To date, 24 MMPs have been identified in mammals. Among these MMPs, some are membrane-type MMPs which are anchored to the cell surface and others are secreted into the extracellular space. In general, MMPs are released as inactive proform MMPs and activated by proteolytic cleavage of the N-terminal domain. In gelatinase subfamily of MMPs (i.e., MMP-2 and MMP-9), the catalytic domain that contains the Zn²⁺ binding site and repeats of fibronectin motifs allowing the ability to bind their major substrate gelatin. MMP-9 (gelatinase B; 92 kDa) is usually low and its expression can be induced by various proinflammatory factors such as cytokines. The other class of gelatinase, MMP-2 (gelatinase A; 72 kDa), is constitutively expressed in several cell types and usually not inducible. In CNS, MMPs, especially, MMP-9 are implicated in several important physiological events, including morphogenesis, wounding healing, and neurite outgrowth [105]. Moreover, upregulation of MMP-9 may contribute to the pathogenesis of several CNS diseases such as stroke, AD, multiple sclerosis, and malignant glioma [105]. Several proinflammatory factors including cytokines, endotoxins, and oxidative stress have been shown to upregulate MMP-9 in astrocytes *in vitro* [106, 107], implying that MMP-9 activity may be regulated by diverse factors in CNS during neuroinflammation. Moreover, many proinflammatory mediators like cytokines and BK induce the expression of MMP-9 during brain injury by increasing ROS production [25, 62]. Recently, upregulated MMP-9 and ROS generation from brain astrocytes have been reported to contribute to neuronal cell death *in vitro* [30]. These studies suggest that upregulation and activation of MMP-9 by proinflammatory factors are mediated through oxidative stress (ROS production) during brain injury and inflammation (Figure 4). Therefore, the inhibition of MMP-9-mediated inflammatory pathways may provide therapeutic strategies to brain inflammation and neurodegenerative diseases.

5.2. Cytosolic Phospholipase A₂. There are three forms of phospholipase A₂ (PLA₂) superfamily including the secretory PLA₂, type IV PLA₂, also known as cPLA₂, and calcium-independent PLA₂ in mammalian cells [108–110]. The secretory PLA₂ (sPLA₂) is expressed in a variety of cell types and it has no preference for AA at *sn*-2 position, requires millimolar amounts of Ca²⁺ for activity and is sensitive to sulfhydryl reducing agents, such as dithiothreitol (DTT), and is resistant to heat or acid conditions [109]. The calcium-independent PLA₂ (iPLA₂) does not require Ca²⁺ for catalytic activity. The iPLA₂ prefers plasmalogen substrates and does not appear to have a preference for the type of fatty acid at the *sn*-2 position. The third class is the novel and high molecular weight (85 kDa) cPLA₂. The cPLA₂ catalyzes the hydrolysis of the *sn*-2 position of membrane glycerophospholipids, leading

to production of free fatty acids and lysophospholipids. This reaction is of particular importance if the esterified fatty acid is AA, which is converted by downstream metabolic enzymes to various bioactive lipophilic compounds called eicosanoids, including PGs and leukotrienes (LTs) [110]. PLA₂ could be the initial and rate-limiting enzyme in this conversion. The increase in cPLA₂ activation and expression following external stimuli, including proinflammatory cytokines, growth factors, and microbial toxin, is often observed in several systems [111]. Among these enzymes, cPLA₂ is the only one that plays a key role in mediating agonist-induced AA release for eicosanoid production in various cell types [112]. Several studies have indicated that cPLA₂ is constitutively expressed in the cytosol of most resting brain cells and tissues. In brain, cPLA₂ has been shown to co-localize with glial fibrillary acidic protein (GFAP), a principal marker for brain astrocytes [113]. Moreover, under brain inflammatory and neurodegenerative conditions such as AD, there is an increase in immunoreactivity to cPLA₂ in astrocytes from the cortex of patients [114, 115]. A variety of proinflammatory factors including IL-1 β , TNF- α , or BK may exert as modulators of cPLA₂ activity and/or expression in various cell types including astrocytes [23, 111]. Upregulation and activation of cPLA₂ leading to PGE₂ production have been implicated in a number of neurodegenerative diseases [111, 114, 115]. Recently, PGE₂ production and cPLA₂ activation have also been shown to regulate the CREB-dependent iNOS expression in microglia [116] or cPLA₂ expression in amnion fibroblasts [117]. However, a series of highly reactive PGs, free fatty acids, lysophospholipids, eicosanoids, platelet-activating factor, and ROS, all generated by enhanced PLA₂ activity and AA release, participate in cellular injury, particularly in neurodegeneration [118]. Thus, cPLA₂ seems to function as a crucial upstream regulator of the production of eicosanoids during brain inflammation and is correlated to the process of neurodegenerative diseases (Figure 4). The inhibition of cPLA₂-mediated pathways may provide a therapeutic strategy to brain inflammation and neurodegenerative diseases.

5.3. Cyclooxygenase-2. COX, known as a prostaglandin-endoperoxide synthase, is a rate-limiting key enzyme in the synthesis of PGs. In this process, PLA₂ catalyzes the release of AA from membrane phospholipids, while COX catalyzes the conversion of AA into PGs [119]. Significant advances have been made in understanding the role of COX in certain biologic processes, including inflammation, angiogenesis, development, and several homeostasis [119]. COX exists in two isoforms: COX-1, which is expressed constitutively under normal conditions in most tissues, mediates regulating normal physiological responses, and controls renal homeostasis, and the inducible COX-2, is not detectable in most normal tissues or resting cells, but its expression can be induced rapidly by a variety of stimuli including cytokines, bacterial or viral infections, and other mediators to produce PGs during inflammation [120]. In addition, COX-2 gene promoter which contains multiple regulatory elements has been shown to be regulated by different transcription factors, including NF- κ B, AP-1, and cyclic AMP-response element binding protein (CREB) in various cell types [121].

Previous studies showed that COX-2 immunoreactivity is a characteristic finding in the synovial macrophage of patients with arthritis as well as in other forms of inflammation. Moreover, several lines of evidence have confirmed COX-2 as a major therapeutic target for the treatment of inflammatory disorders such as arthritis [119, 122]. Recently, the mice with homozygous deletion of the COX-2 gene suppress endotoxin-induced inflammation [123]. In brain, expression of COX-2 leads to increased production of prostanoids which are potent inflammatory mediators, and upregulated COX-2 expression has been reported in neurodegenerative disorders [124]. Moreover, upregulation of COX-2 and PGE₂ release by viral infection such as EV71 have been reported in brain astrocytes and human neuroblastoma cells via diverse signaling pathways [125, 126]. Upregulation of COX-2/PGE₂ by ET-1 via MAPK-dependent NF- κ B pathway in brain microvascular endothelial cells [127]. A recent report also indicates that the ROS-induced COX-2 expression can be found in ALS [128]. However, the expression of COX-2 appears to be strongly induced and activated during AD, indicating the importance of inflammatory gene pathways as a response to brain injury [118]. Thus, COX-2 may play an important role in the development of brain inflammation and neurodegenerative diseases.

5.4. Nitric Oxide Synthase. NO is a free radical that displays diverse bioactivity in various organ systems, including CNS. Depending on the concentration, excess NO levels are implicated in the pathogenesis of CNS diseases including ischemia, trauma, neuroinflammatory, and neurodegenerative diseases [129–131]. Production of NO from L-arginine is catalyzed by NOS. The level of iNOS in healthy brain is undetectable. Accumulating evidence supports the role of iNOS in the pathogenesis of CNS disorders. In CNS, upregulation of iNOS in various cell types, including astrocytes and microglia, is proposed to be the leading source of NO production during neuroinflammation [132]. Furthermore, knockout strategies of iNOS gene protect against focal cerebral ischemia and LPS challenges [133, 134]. iNOS is induced by a variety of stimuli, such as viral and bacterial infections, cytokines, cell-cell contact, and neurotoxins [131]. The consequent product NO reacts with superoxide to form peroxynitrite (ONOO⁻), the most toxic derivative of NO (Figure 3). As for the involvement of NO derivatives in neuropathology, many studies have revealed that the reference of iNOS/NO/ONOO⁻ plays an important role in neurodegenerative disorders [131]. However, following inflammatory insults, reactive astrocytes express iNOS, which causes the neuronal damage associated with cerebral ischemia and/or demyelinating diseases [132]. In CNS, appearance of iNOS in astrocytes is related to several neurodegenerative diseases such as ALS [130] and multiple sclerosis (MS) [129]. These findings imply that astrocytes are the leading regulators in neurodegenerative diseases. Moreover, activation of astrocytes has been reported to involve in the expression of inflammatory genes. It has been well established that the regulation of iNOS expression is mediated via tyrosine kinases such as JAK, MAPKs, ROS, and various transcription factors including STAT-1, NF- κ B, and AP-1 in astrocytes [131]. Increasing evidence suggests

that activation of signal transduction pathways like c-Src, PI3K/Akt, and MAPK cascades contributes to activation of astrocytes and microglia, leading to expression of inflammatory proteins and advanced damage in neurodegenerative diseases [25, 26, 135].

5.5. Adhesion Molecules. Cell adhesion molecules play an important role in inflammatory responses. Leukocytes continuously circulate throughout the body in order to come in contact with antigens sequestered within tissues. To enter tissues, circulating leukocytes migrate from the blood between vascular endothelial cells and into the tissue [136]. During this migration, leukocytes initially bind to endothelial cells via low-affinity adhesion molecules. The low-affinity adhesion in combination with the force of the blood flow results in rolling leukocytes on endothelial cells. Subsequently, adhesion molecule affinity is upregulated and leukocytes firmly adhere to the endothelium [136]. Finally, bound leukocytes migrate between the endothelial cells and into the tissue. The vascular cell adhesion molecule 1 (VCAM-1) is one of the inducible cell transmembrane glycoproteins of the immunoglobulin supergene family expressed on several cell types and plays an important role in a number of inflammatory and immune responses [137]. It was first identified as an adhesion molecule induced on endothelial cells by proinflammatory cytokines or LPS [138]. VCAM-1 expression is induced on endothelial cells during inflammatory bowel disease, atherosclerosis, and infections [139]. Upregulation of VCAM-1 expression on cytokine-triggered vascular endothelial cells enhances the targeted transmigration of PMNs into extravascular space of inflammation [137]. In brain, proinflammatory cytokine-mediated expression of cell surface adhesion molecules plays a key role in endothelial cell injury, leading to vascular inflammation and the development of many cerebrovascular diseases [140]. Moreover, astrocytes can be induced by viral infections to express the adhesion molecules. Upregulation of adhesion molecules such as ICAM-1 (intercellular adhesion molecule 1) and VCAM-1 in astrocytes is required for monocyte-astrocyte interaction which increases infiltration of monocytes into the CNS observed in the patients with HIV-1 dementia [141]. HIV-1 Tat enhances monocyte adhesion by upregulation of ICAM-1 and VCAM-1 genes via a ROS-dependent NF- κ B activation in astrocytes [141]. Understanding the role of ROS in proinflammatory factor-mediated adhesion molecule expression and subsequently increased adhesion of monocyte to brain cells provides an occasion for the development of anti-inflammatory compounds that may be useful as therapeutic strategies for the CNS inflammation and ROS-associated neurotoxicity.

5.6. Stress Protective Proteins. In contrast with inflammatory proteins, recent reports indicate that the ROS can also induce several stress protective proteins, such as HO-1 and heat-shock proteins (HSP70 in particular), which may exert protective effects from the deleterious effects of inflammation [142]. Abnormal protein folding has been shown as a cause of various diseases like neurodegenerative diseases in

association with inflammatory mechanisms. In the events, the HSPs play a crucial role in preventing protein misfolding and inhibiting apoptotic activity and represent a class of proteins potentially involved in PD pathogenesis [143]. Recent studies have shown that HSPs are colocalized in protein aggregates in AD, PD, and other neurodegenerative disorders [144, 145]. Many experimental findings have demonstrated that selective overexpression of HSP70 prevents the disease progression in various animal models and cellular models [145]. Furthermore, HSP70 dysfunction activates intracellular signaling like NF- κ B that can also promote neurodegeneration [146]. Thus, the expression of HSP70 may prove diagnostic and prognostic values in inflammatory conditions and therapeutical applications are being considered on the basis of these reports.

6. Redox Signal-Mediated Signaling Transduction

Recently, increasing evidence has demonstrated that oxidative stress (ROS generation) also plays a key signaling molecule in regulation of various inflammatory mediators in several cell types. Although many cells from brain tissue can produce various inflammatory mediators [42, 105], the intracellular signaling mechanisms responsible for the regulation of diverse inflammation-relating mediators expression induced by proinflammatory factors in brain cells like astrocytes are not completely characterized. Next, we review some signaling molecules in several inflammatory target protein expressions induced by proinflammatory factors in brain cells.

6.1. Mitogen-Activated Protein Kinases. Many proinflammatory cytokines and chemokines transducer signals are mediated via activation of MAPKs pathways. There is growing evidence that members of the MAPK family may play a central role in neurodegeneration [147]. MAPKs are important components of signaling modules activated by neurotransmitters, cytokines, and growth factors, as well as chemical and mechanical stressors. In mammals, three groups of MAPKs have been identified: the extracellular signal-regulated protein kinases (ERKs), the c-Jun NH₂-terminal kinases (JNKs), and the p38 MAPK. ERK is activated by diverse stimuli, including growth factors and cytokines [147]. The p38 MAPK is activated by cellular stresses, including cytokines, LPS, growth factors, and UV radiation. The JNK is activated by many of the same stimuli that activate p38 MAPK, such as cellular stresses and various cytokines. Moreover, abnormal MAPK regulation might be implicated in CNS injury and inflammation [148]. Several mediators such as BK have been reported to act as an important proinflammatory factors through activation of MAPK cascades in different cell types [21–26]. In brain cells, the activation of ERK1/2 is mainly associated with proliferation, differentiation, and development in response to nerve growth factors. In contrast, the JNK and p38 MAPK signaling pathways are activated by various environmental stress and inflammatory factors that have been shown to promote neuronal cell death [149].

Moreover, the JNK and p38 MAPK signaling cascades can also be strongly activated by stress-induced ROS production or a mild oxidative shift of the redox state [28]. Both JNK and p38 MAPK are recognized as contributors to neurodegeneration by their ability to mediate intracellular stress events in transgenic mouse models of AD [19]. The p38 MAPK activation and COX-2 and PGE₂ induction are served as contributors to neuronal damage in AD in response to oxidative stress [150].

In nonneural cells like astrocytes, many studies have found that A β peptide can activate astrocytes, including morphological alterations, cytokine induction, NO release [151], and chemokine and matrix-degrading proteinases production [152]. These findings further indicate that induction of several inflammatory mediators by the A β -stimulated activation of MAPKs in glial cells may be involved in AD progression. Moreover, our recent reports in astrocytes have demonstrated that the proinflammatory factors including TGF- β and BK can induce many inflammatory mediators such as MMP-9 expression through the ROS-dependent MAPK cascades [40]. These results suggest that upregulation of inflammatory mediators via ROS-mediated activation of MAPKs in astrocytes might play a key role during the CNS inflammation and neurodegeneration. Moreover, these results also implicate that the distinct groups of MAPKs are activated by a ROS-dependent manner which contribute to the expression of various inflammatory genes and are dependent on the external stimuli during brain inflammation. Thus, ROS may mediate MAPKs activation and expression of inflammatory genes in response to proinflammatory mediators in the CNS inflammatory disorders (Figure 5).

6.2. Transactivation of Receptor Tyrosine Kinases. Cross-communication between different signaling systems allows the integration of the great diversity of stimuli that a cell receives under varying physiological situations. The most direct mechanism is receptor heterodimerization that is well described for members of the epidermal growth factor receptor (EGFR) family [153]. In addition to growth factor receptor tyrosine kinases (RTKs) cross-talk, also completely unrelated cell surface receptors are able to communicate and influence each other, which play a key role in the transmission of information from outside the cell into the cytoplasm and nucleus. A variety of cytokines and growth factors that act as respective receptors have been reported to induce production of ROS in nonimmune cells. The prototype for such a pathway is the GPCR-induced transactivation of EGFR signal [154]. Treatment of cells with GPCR agonists induces phosphorylation of the EGFR by metalloprotease-dependent release of EGF-like ligands such as HB-EGF, thereby coupling GPCRs to EGFR characteristic downstream signaling pathways such as MAPKs or PI3K/Akt pathway [155]. In addition to the EGFR, other RTKs have been shown to be activated in response to GPCR stimulation, comprising the Trk receptor [156] and platelet-derived growth factor receptor (PDGFR) [157]. Previous studies have shown that in developing carcinoma cells, the early effects of COX-2-derived PGE₂ and lysophosphatidic acid are in part mediated by the EGFR or PDGER, and this transactivation is responsible for

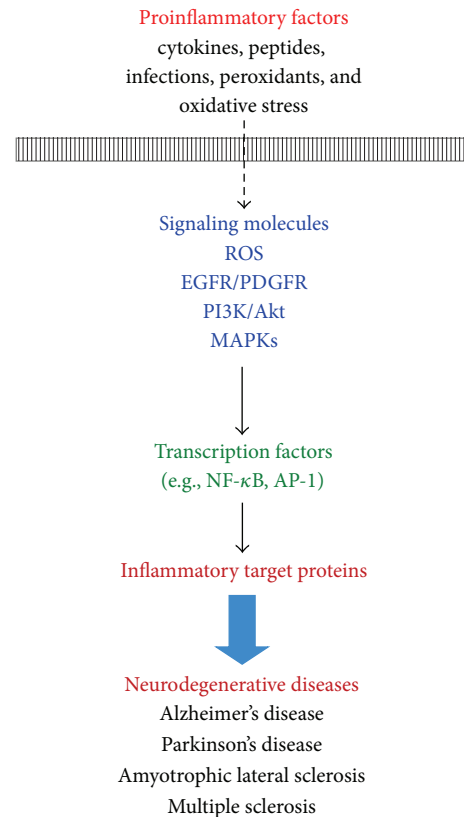


FIGURE 5: Proposed mechanisms of proinflammatory factors-stimulated activation of various signaling molecules and transcription factors leading to the expression of inflammatory target genes in brain resident cells. The intracellular signaling molecules include ROS, EGFR/PDFER, PI3K/Akt, and MAPKs. Oxidative stress may regulate these signaling pathways leading to activation of transcription factors such as NF- κ B and AP-1 and recruitment of coactivator p300 in the transcription initiation complex. Ultimately, upregulation of diverse inflammatory target proteins can cause the pathogenesis of several neurodegenerative diseases. EGFR: epidermal growth factor receptor; PDGFR: platelet-derived growth factor receptor; PI3K: phosphoinositide-3'-kinase; MAPKs: mitogen-activated protein kinases; NF- κ B: Nuclear factor- κ B; AP-1: activator protein-1.

subsequent downstream effects including the stimulation of cell migration and invasion [158]. However, receptor cross-talk can also occur in a ligand-independent manner involving for instance, non-RTKs such as c-Src [159]. Production of ROS results from the activation of signaling through the EGF and PDGF receptors [160]. In addition, ROS have been shown to stimulate c-Src-dependent transactivation of PDGFR α [161]. Accumulating evidence has shown that PKC-dependent activation of Nox is essential for PDGF-stimulated ROS generation, which is important for PDGF-induced MAPKs activation [162]. In the adult CNS, the EGFR pathway is highly upregulated and activated in astrocytes following neuronal injury [163]. Activation of the EGFR pathway triggers quiescent astrocytes to become reactive astrocytes that appear to be destructive to neurons in the adult CNS [163]. Regulation of RTKs such as EGFR in

astrocytes may be a new therapeutic strategy for the treatment of neural disorders. These studies suggest that growth factor RTKs may play a pivotal role in mediating inflammatory genes regulation through ROS signal in several diseases including the CNS disorders (Figure 5).

6.3. Phosphoinositide-3'-Kinase (PI3K)/Akt Cascade. The phosphoinositide-3'-kinase (PI3K)/Akt cascade, the common downstream signal of EGF and PDGF receptors, is a cell survival pathway and regulated by various growth factor receptor-dependent mechanisms. Recent studies suggested that numerous components of the PI3K/Akt pathway play a crucial role in the expression and activation of inflammatory mediators, inflammatory cell recruitment, immune cell function, and tissue remodeling in chronic inflammatory diseases. In astrocytes, we demonstrated that ET-1 induced iNOS expression and NO production through PI3K/Akt cascade [26]. Moreover, PI3K/Akt cascade contributes to the expression of various inflammatory mediators induced by several proinflammatory factors in brain cells including astrocytes [125, 127]. Selective PI3K inhibitors such as wortmannin and LY294002 have been developed that reduce inflammation and some characteristics of disease in experimental animal models. In addition, ROS induction is often accompanied by the activation of PI3K/Akt cascade. For example, LY294002 has been shown to reduce chemokine-induced ROS generation in phagocytes [164], which was further confirmed by studies using PI3K knockout mice. Many studies have indicated the ROS generation induced by cytokines, PDGF, or VEGF in several cell types, which is reduced by inhibition of PI3K activity, suggesting that PI3K is involved in the ROS production induced by cytokines and growth factors. In addition to the role of PI3K/Akt cascade in ROS production, several reports support that the opposite hierarchical relationship exists between ROS and PI3K/Akt cascade. PI3K/Akt was activated in response to the exogenous treatment of H₂O₂ in several cell types [165]. Moreover, ROS have been shown to regulate phosphorylation of Akt [166] and then induce the expression of inflammatory genes associated with inflammation in various cell types. Taken together, these results implicate that ROS-dependent PI3K/Akt cascade or PI3K/Akt-mediated ROS signal may be critical for regulating the expression of inflammatory proteins in the brain inflammation and neurodegenerative disorders (Figure 5).

6.4. Transcription Factors. The progressive increase of oxidative stress during injuries not only causes oxidative damage to cellular macromolecules, but also modulates the pattern of gene expression through functional alterations of transcription factors. Here we focus on the roles of many transcription factors (e.g., NF- κ B and AP-1), which are well known to be modulated during oxidative stress associated with physiological and pathological events [32]. The transcription factors such as NF- κ B and AP-1 play a key role in the regulation of several gene expressions including proinflammatory cytokines, adhesion molecules, chemokines, growth factors, and inducible enzymes (e.g., MMPs, cPLA₂, COX-2, and iNOS) during inflammation, immunity, cell proliferation,

stress response, and apoptosis [167–169]. One important and widely investigated transcription factor which is NF- κ B is a major participant in signaling pathways governing cellular responses to environmental (oxidative) stresses [168]. The nuclear translocation and activation of NF- κ B in response to various stimuli, such as proinflammatory cytokines, LPS, and oxidative challenge (ROS production), are sequentially organized at the molecular level [168]. Moreover, NF- κ B act as a positive regulator in the expression of many inflammatory genes such as COX-2 involved in chronic inflammatory diseases [169]. Cytokines such as IL-1 β and TNF- α have been shown to activate NF- κ B leading to upregulation of various NF- κ B-dependent genes in several cell types [168]. It is of interest that many of the genes regulated by these MAPK pathways are dependent on NF- κ B for transcription and lead to expression of inflammatory genes such as MMP-9 at the transcriptional level [169, 170]. In astrocytes, various stimuli can induce the expression of several inflammatory mediators, including MMP-9, cPLA₂, COX-2, and iNOS, through ROS-mediated activation of NF- κ B manner [40, 62].

In addition, activator protein-1 (AP-1) is a sequence-specific transcriptional activator mainly composed of members of the Fos, Jun, and ATF-2 families. These proteins associate to form a variety of homodimers or heterodimers that bind to an AP-1 binding element within the promoter region of inflammatory genes such as COX-2 and MMP-9. It is a well-known redox-regulated transcription factor for the expression of several AP-1-dependent genes induced by diverse stress signals such as ROS generation associated with physiological and pathological events [25, 62, 170]. Several reports indicate that AP-1 is also involved in the pathogenesis of brain inflammation (Figure 5). Many studies have demonstrated that ROS signals (e.g., O₂^{•-} and H₂O₂) contribute to the expression or activation of AP-1 proteins (e.g., c-Fos) [62]. Recently, Kim et al. demonstrated that apocynin (a Nox inhibitor) shows potential antioxidant activities and inhibitory effects on the activation of redox-sensitive transcription factors, such as AP-1, induced by proinflammatory stimuli such as TNF- α [171]. The reports indicate that CSE induces cPLA₂ expression through the production of ROS and subsequent activation of the MAPK pathway and AP-1 in human tracheal smooth muscle cells [172]. In astrocytes, we have demonstrated that AP-1 participates in the expression of several genes, including MMP-9 and HO-1, by BK through ROS-dependent manner [25, 62]. These results implicate that ROS play a central role in regulating AP-1 activation or expression and lead to inflammatory genes expression in brain inflammation and neurodegenerative disorders (Figure 5).

6.5. Transcription Coactivators. The transcription coactivator p300/CREB binding protein (CBP) is vital for the coactivation of several transcription factors such as NF- κ B and AP-1 in the transcription machinery, which has a significant role in the activation of transcription factor-mediated gene expression for proinflammatory factors [173–175]. The p300 protein is a key regulator of RNA polymerase II-mediated transcription. Several studies indicate that p300 participates in the expression of inflammatory genes induced by cytokines

and growth factors. Furthermore, the transcriptional cofactor p300/CBP is an important component of the transcriptional machinery that participates in regulation at the levels of both chromatin modification and transcription initiation [173–175]. Previous studies have indicated that the promoter of several gene transcriptions, chromatin remodeling, and histone modification is regulated by p300/CBP [175]. However, in astrocytes, the p300 is vital for the coactivation of several transcription factors such as AP-1 in the transcription machinery, which has a significant role in the activation of AP-1-mediated gene expression for proinflammatory mediators [173]. Previous results have indicated that p300 plays an important role in BK-, IL-1 β -, and oxLDL-induced MMP-9 expression in astrocytes [21, 22, 96]. Recently, a study has shown that ROS-dependent p300 activation leads to cPLA₂ expression by cigarette smoke extract in human tracheal smooth muscle cells [172]. Consistently, we have demonstrated that LTA induces p300/AP-1-dependent MMP-9 expression via ROS-mediated pathway in astrocytes [27]. Moreover, oxidative stress activates NF- κ B resulting in the expression of proinflammatory mediators through the activation of intrinsic HAT activity on coactivator molecules. Oxidative stress also inhibits HDAC activity and in doing so enhances the expression of inflammatory genes which leads to a chronic inflammatory response. Oxidative stress can also increase complex formation between the coactivator p300 and the p65 subunit of NF- κ B suggesting a further role of oxidative stress in chromatin remodeling [1]. Together, these studies indicate that the oxidative stress-stimulated coactivator p300 may play a critical role in the expression of inflammatory genes during brain inflammation and neurodegenerative disorders.

7. Conclusions

Glial cells maintain brain plasticity and protect the brain for functional recovery from injuries. Reactivation of glial cells may promote neuroinflammation and neurodegeneration (Figure 1) and, ultimately, the retraction of neuronal synapses, which leads to cognitive deficits [10]. Moreover, redox signaling is a critical event in several inflammatory diseases such as AD that precedes the formation of these disease pathologies. To date, although numerous effects have been made to develop therapies based on antioxidants in the past years, the actual benefits to the patients have been very limited. It is likely due to lack of potency, late administration, and poor penetration into the brain cells [7, 32]. Alternative strategies including searching for factors that initiate endogenous antioxidants are necessary to improve the efficacy of treatment (Figure 2). Moreover, increased oxidative stresses (ROS) by various proinflammatory factors, such as cytokines, peptides, bacterial or viral infections, peroxidants, and other stress, serve as intracellular signals in gene regulation and signaling transduction, in addition to their deleterious effects on cellular components. Thus, understanding how oxidative stress produces and modulates expression of several genes that might help to develop effective therapeutic strategies for CNS diseases. First, the focus

of this review is on glial cells and their effects on the CNS disorders. Moreover, this review summarized the interplay between oxidative stress and neuroinflammation via ROS production which contributes to neurodegeneration, thereby enhancing disease progression based on data collected from brain cells, particularly astrocytes, in *in vitro* and *in vivo* studies (Figure 1). Perhaps modifying the activity of glial cells to reduce their neurotoxic properties and enhance their neuroprotective effects may offer potential targets for therapeutic interventions in neurodegenerative diseases. Oxidative stress-induced signaling transduction pathways, including ROS, transactivation of EGFR or PDGFR, PI3K/Akt, MAPKs, NF- κ B, and AP-1, that are associated with the CNS disorders were discussed (Figure 4). Moreover, the review highlighted current progress on the association of oxidative stress with the expression of various inflammatory genes, including MMP-9, cPLA₂, COX-2, iNOS, and adhesion molecules and redox signal-sensitive transcription factors that may contribute to the development of the CNS inflammation and neurodegenerative diseases (Figure 5). Possible therapeutic strategies to target redox-sensitive signaling molecules, transcription factors, or cofactors are implicated based on the updated view of ROS-mediated regulation of inflammatory target genes in brain inflammation and neurodegenerative disorders.

Abbreviations

ROS:	Reactive oxygen species
CNS:	Central nervous system
AD:	Alzheimer's disease
PD:	Parkinson's disease
MMPs:	Matrix metalloproteinases
cPLA ₂ :	Cytosolic phospholipase A ₂
COX-2:	Cyclooxygenase-2
Nox2:	NADPH oxidase 2
iNOS:	Inducible nitric oxide synthase
LPS:	Lipopolysaccharide
IL-1 β :	Interleukin-1
TNF- α :	Tumor necrosis factor- α
BBB:	Blood-brain barrier
TLRs:	Toll-like receptors
PGs:	Prostaglandins
NO:	Nitric oxide
A β :	β -Amyloid
BK:	Bradykinin
ET-1:	Endothelin-1
oxLDL:	Oxidized low-density lipoprotein
HO-1:	Heme oxygenase-1
CO:	Carbon monoxide
RNS:	Reactive nitrogen species
Xox:	Xanthine oxidase
GPCR:	G-Protein-coupled receptor
LTA:	Lipoteichoic acid
JEV:	Japanese encephalitis virus
EV71:	Enterovirus 71
AA:	Arachidonic acid
VCAM-1:	Vascular cell adhesion molecule 1
MAPKs:	Mitogen-activated protein kinases
ERKs:	Extracellular signal-regulated protein kinases

JNKs: c-Jun NH₂-terminal kinases
 EGFR: Epidermal growth factor receptor
 RTKs: Receptor tyrosine kinases
 PDGFR: Platelet-derived growth factor receptor
 PI3K: Phosphoinositide-3'-kinase
 NF- κ B: Nuclear factor- κ B
 AP-1: Activator protein 1
 CREB: Cyclic AMP-response element binding protein
 CBP: CREB binding protein.

Conflict of Interests

The authors declare that they have no conflict of interests.

Acknowledgments

This work was supported by National Science Council, Taiwan, Grant nos. NSC102-2321-B-182-011, NSC101-2320-B-182-039-MY3, and NSC102-2320-B-255-005-MY3; Chang Gung Medical Research Foundation, Grant nos. CMRPD1C0101, CMRPD1B0382, CMRPD1C0561, CMRPF1C0191, and CMRPF1A0063; and the Ministry of Education, Taiwan; Grant nos. EMRPD1C0261 and EMRPD1C0271.

References

- I. Rahman, J. Marwick, and P. Kirkham, "Redox modulation of chromatin remodeling: impact on histone acetylation and deacetylation, NF- κ B and pro-inflammatory gene expression," *Biochemical Pharmacology*, vol. 68, no. 6, pp. 1255–1267, 2004.
- M. Valko, D. Leibfritz, J. Moncol, M. T. D. Cronin, M. Mazur, and J. Telser, "Free radicals and antioxidants in normal physiological functions and human disease," *International Journal of Biochemistry and Cell Biology*, vol. 39, no. 1, pp. 44–84, 2007.
- I. T. Lee and C. M. Yang, "Role of NADPH oxidase/ROS in pro-inflammatory mediators-induced airway and pulmonary diseases," *Biochemical Pharmacology*, vol. 84, no. 5, pp. 581–590, 2012.
- W. Dröge, "Free radicals in the physiological control of cell function," *Physiological Reviews*, vol. 82, no. 1, pp. 47–95, 2002.
- R. von Bernhardi and J. Eugenin, "Alzheimer's disease: redox dysregulation as a common denominator for diverse pathogenic mechanisms," *Antioxidants and Redox Signaling*, vol. 16, no. 9, pp. 974–1031, 2012.
- B. Halliwell, "Oxidative stress and neurodegeneration: where are we now?" *Journal of Neurochemistry*, vol. 97, no. 6, pp. 1634–1658, 2006.
- B. Uttara, A. V. Singh, P. Zamboni, and R. T. Mahajan, "Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options," *Current Neuropharmacology*, vol. 7, no. 1, pp. 65–74, 2009.
- A. Melo, L. Monteiro, R. M. F. Lima, D. M. de Oliveira, M. D. de Cerqueira, and R. S. El-Bachá, "Oxidative stress in neurodegenerative diseases: mechanisms and therapeutic perspectives," *Oxidative Medicine and Cellular Longevity*, vol. 2011, Article ID 467180, 14 pages, 2011.
- V. Chiurchiù and M. MacCarrone, "Chronic inflammatory disorders and their redox control: from molecular mechanisms to therapeutic opportunities," *Antioxidants and Redox Signaling*, vol. 15, no. 9, pp. 2605–2641, 2011.
- D. Farfara, V. Lifshitz, and D. Frenkel, "Neuroprotective and neurotoxic properties of glial cells in the pathogenesis of Alzheimer's disease: Alzheimer's review series," *Journal of Cellular and Molecular Medicine*, vol. 12, no. 3, pp. 762–780, 2008.
- S. Fuller, M. Steele, and G. Münch, "Activated astroglia during chronic inflammation in Alzheimer's disease-Do they neglect their neurosupportive roles?" *Mutation Research*, vol. 690, no. 1-2, pp. 40–49, 2010.
- H. K. Kimelberg, "Receptors on astrocytes—what possible functions?" *Neurochemistry International*, vol. 26, no. 1, pp. 27–40, 1995.
- L. F. Eng and R. S. Ghirnikar, "GFAP and astrogliosis," *Brain Pathology*, vol. 4, no. 3, pp. 229–237, 1994.
- Y. S. Kim and T. H. Joh, "Microglia, major player in the brain inflammation: their roles in the pathogenesis of Parkinson's disease," *Experimental and Molecular Medicine*, vol. 38, no. 4, pp. 333–347, 2006.
- M. Eddelston and L. Mucke, "Molecular profile of reactive astrocytes—implications for their role in neurologic disease," *Neuroscience*, vol. 54, no. 1, pp. 15–36, 1993.
- J. L. Ridet, S. K. Malhotra, A. Privat, and F. H. Gage, "Reactive astrocytes: cellular and molecular cues to biological function," *Trends in Neurosciences*, vol. 20, no. 12, pp. 570–577, 1997.
- G. C. Brown, "Mechanisms of inflammatory neurodegeneration: INOS and NADPH oxidase," *Biochemical Society Transactions*, vol. 35, no. 5, pp. 1119–1121, 2007.
- M. Koistinaho, M. I. Kettunen, G. Goldsteins et al., " β -amyloid precursor protein transgenic mice that harbor diffuse A β deposits but do not form plaques show increased ischemic vulnerability: role of inflammation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 3, pp. 1610–1615, 2002.
- M. J. Savage, Y.-G. Lin, J. R. Ciallella, D. G. Flood, and R. W. Scott, "Activation of c-Jun N-Terminal Kinase and p38 in an Alzheimer's Disease Model Is Associated with Amyloid Deposition," *The Journal of Neuroscience*, vol. 22, no. 9, pp. 3376–3385, 2002.
- R. E. Mrak, J. G. Sheng, and W. S. T. Griffin, "Glial cytokines in Alzheimer's disease: review and pathogenic implications," *Human Pathology*, vol. 26, no. 8, pp. 816–823, 1995.
- C.-Y. Wu, H.-L. Hsieh, C.-C. Sun, C.-P. Tseng, and C.-M. Yang, "IL-1 β induces proMMP-9 expression via c-Src-dependent PDGFR/PI3K/Akt/p300 cascade in rat brain astrocytes," *Journal of Neurochemistry*, vol. 105, no. 4, pp. 1499–1512, 2008.
- H.-L. Hsieh, C.-Y. Wu, and C.-M. Yang, "Bradykinin induces matrix metalloproteinase-9 expression and cell migration through a PKC- δ -dependent ERK/Elk-1 pathway in astrocytes," *Glia*, vol. 56, no. 6, pp. 619–632, 2008.
- H.-L. Hsieh, C.-Y. Wu, T.-L. Hwang, M.-H. Yen, P. Parker, and C.-M. Yang, "BK-induced cytosolic phospholipase A2 expression via sequential PKC- δ , p42/p44 MARK, and NF- κ B activation in rat brain astrocytes," *Journal of Cellular Physiology*, vol. 206, no. 1, pp. 246–254, 2006.
- H.-L. Hsieh, H.-H. Wang, C.-Y. Wu et al., "BK-induced COX-2 expression via PKC- δ -dependent activation of p42/p44 MAPK and NF- κ B in astrocytes," *Cellular Signalling*, vol. 19, no. 2, pp. 330–340, 2007.
- C. C. Lin, H. L. Hsieh, R. H. Shih et al., "NADPH oxidase 2-derived reactive oxygen species signal contributes

- to bradykinin-induced matrix metalloproteinase-9 expression and cell migration in brain astrocytes," *Cell Communication and Signaling*, vol. 10, no. 1, p. 35, 2012.
- [26] H.-H. Wang, H.-L. Hsieh, and C.-M. Yang, "Nitric oxide production by endothelin-1 enhances astrocytic migration via the tyrosine nitration of matrix metalloproteinase-9," *Journal of Cellular Physiology*, vol. 226, no. 9, pp. 2244–2256, 2011.
- [27] H. L. Hsieh, C. C. Lin, R. H. Shih, L. D. Hsiao, and C. M. Yang, "NADPH oxidase-mediated redox signal contributes to lipoteichoic acid-induced MMP-9 upregulation in brain astrocytes," *Journal of Neuroinflammation*, vol. 9, p. 110, 2012.
- [28] W.-H. Tung, H.-W. Tsai, I.-T. Lee et al., "Japanese encephalitis virus induces matrix metalloproteinase-9 in rat brain astrocytes via NF-KB signalling dependent on MAPKs and reactive oxygen species," *British Journal of Pharmacology*, vol. 161, no. 7, pp. 1566–1583, 2010.
- [29] H.-H. Wang, H.-L. Hsieh, C.-Y. Wu, C.-C. Sun, and C.-M. Yang, "Oxidized low-density lipoprotein induces matrix metalloproteinase-9 expression via a p42/p44 and JNK-dependent AP-1 pathway in brain astrocytes," *Glia*, vol. 57, no. 1, pp. 24–38, 2009.
- [30] C. M. Yang, H. L. Hsieh, C. C. Lin et al., "Multiple factors from bradykinin-challenged astrocytes contribute to the neuronal apoptosis: involvement of astroglial ROS, MMP-9, and HO-1/CO system," *Molecular Neurobiology*, vol. 47, no. 3, pp. 1020–1033, 2013.
- [31] S. Chrissobolis and F. M. Faraci, "The role of oxidative stress and NADPH oxidase in cerebrovascular disease," *Trends in Molecular Medicine*, vol. 14, no. 11, pp. 495–502, 2008.
- [32] Q. Shi and G. E. Gibson, "Oxidative stress and transcriptional regulation in Alzheimer disease," *Alzheimer Disease and Associated Disorders*, vol. 21, no. 4, pp. 276–291, 2007.
- [33] I. T. Demchenko, T. D. Oury, J. D. Crapo, and C. A. Piantadosi, "Regulation of the brain's vascular responses to oxygen," *Circulation Research*, vol. 91, no. 11, pp. 1031–1037, 2002.
- [34] P. H. Chan, "Reactive oxygen radicals in signaling and damage in the ischemic brain," *Journal of Cerebral Blood Flow and Metabolism*, vol. 21, no. 1, pp. 2–14, 2001.
- [35] F. Serrano and E. Klann, "Reactive oxygen species and synaptic plasticity in the aging hippocampus," *Ageing Research Reviews*, vol. 3, no. 4, pp. 431–443, 2004.
- [36] H. Kamata and H. Hirata, "Redox regulation of cellular signalling," *Cellular Signalling*, vol. 11, no. 1, pp. 1–14, 1999.
- [37] A. Federico, E. Cardaioli, P. da Pozzo, P. Formichi, G. N. Gallus, and E. Radi, "Mitochondria, oxidative stress and neurodegeneration," *Journal of the Neurological Sciences*, vol. 322, no. 1-2, pp. 254–262, 2012.
- [38] J. Kang, E. J. Park, I. Jou, J.-H. Kim, and E.-H. Joe, "Reactive oxygen species mediate A β (25-35)-induced activation of BV-2 microglia," *NeuroReport*, vol. 12, no. 7, pp. 1449–1452, 2001.
- [39] L. Qin, Y. Liu, T. Wang et al., "NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia," *The Journal of Biological Chemistry*, vol. 279, no. 2, pp. 1415–1421, 2004.
- [40] H.-L. Hsieh, H.-H. Wang, W.-B. Wu, P.-J. Chu, and C.-M. Yang, "Transforming growth factor- β 1 induces matrix metalloproteinase-9 and cell migration in astrocytes: roles of ROS-dependent ERK- and JNK-NF- κ B pathways," *Journal of Neuroinflammation*, vol. 7, article 88, 2010.
- [41] P. L. McGeer and E. G. McGeer, "The inflammatory response system of brain: implications for therapy of Alzheimer and other neurodegenerative diseases," *Brain Research Reviews*, vol. 21, no. 2, pp. 195–218, 1995.
- [42] G. A. Rosenberg, "Matrix metalloproteinases in neuroinflammation," *Glia*, vol. 39, no. 3, pp. 279–291, 2002.
- [43] H. Fillit, W. Ding, L. Buee et al., "Elevated circulating tumor necrosis factor levels in Alzheimer's disease," *Neuroscience Letters*, vol. 129, no. 2, pp. 318–320, 1991.
- [44] S. M. Allan, P. J. Tyrrell, and N. J. Rothwell, "Interleukin-1 and neuronal injury," *Nature Reviews Immunology*, vol. 5, no. 8, pp. 629–640, 2005.
- [45] K. Fassbender, S. Rossol, T. Kammer et al., "Proinflammatory cytokines in serum of patients with acute cerebral ischemia: kinetics of secretion and relation to the extent of brain damage and outcome of disease," *Journal of the Neurological Sciences*, vol. 122, no. 2, pp. 135–139, 1994.
- [46] J. A. Smith, A. Das, S. K. Ray, and N. L. Banik, "Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases," *Brain Research Bulletin*, vol. 87, no. 1, pp. 10–20, 2012.
- [47] D. A. Siwik and W. S. Colucci, "Regulation of matrix metalloproteinases by cytokines and reactive oxygen/nitrogen species in the myocardium," *Heart Failure Reviews*, vol. 9, no. 1, pp. 43–51, 2004.
- [48] P. Thornton, E. Pinteaux, R. M. Gibson, S. M. Allan, and N. J. Rothwell, "Interleukin-1-induced neurotoxicity is mediated by glia and requires caspase activation and free radical release," *Journal of Neurochemistry*, vol. 98, no. 1, pp. 258–266, 2006.
- [49] N. H. Greig, M. P. Mattson, T. Perry et al., "New therapeutic strategies and drug candidates for neurodegenerative diseases: p53 and TNF- α inhibitors, and GLP-1 receptor agonists," *Annals of the New York Academy of Sciences*, vol. 1035, pp. 290–315, 2004.
- [50] D. A. Butterfield, J. Drake, C. Pocernich, and A. Castegna, "Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid β -peptide," *Trends in Molecular Medicine*, vol. 7, no. 12, pp. 548–554, 2001.
- [51] C. Caspersen, N. Wang, J. Yao et al., "Mitochondrial A β : a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease," *The FASEB Journal*, vol. 19, no. 14, pp. 2040–2041, 2005.
- [52] V. Della Bianca, S. Dusi, E. Bianchini, I. Dal Prà, and F. Rossi, " β -amyloid activates the O $_2^-$ forming NADPH oxidase in microglia, monocytes, and neutrophils. A possible inflammatory mechanism of neuronal damage in Alzheimer's disease," *The Journal of Biological Chemistry*, vol. 274, no. 22, pp. 15493–15499, 1999.
- [53] G. P. Lim, T. Chu, F. Yang, W. Beech, S. A. Frautschy, and G. M. Cole, "The curry spice curcumin reduces oxidative damage and amyloid pathology in an Alzheimer transgenic mouse," *The Journal of Neuroscience*, vol. 21, no. 21, pp. 8370–8377, 2001.
- [54] Q. Ding, E. Dimayuga, and J. N. Keller, "Oxidative damage, protein synthesis, and protein degradation in Alzheimer's disease," *Current Alzheimer Research*, vol. 4, no. 1, pp. 73–79, 2007.
- [55] H. M. Schipper, D. A. Bennett, A. Liberman et al., "Glial heme oxygenase-1 expression in Alzheimer disease and mild cognitive impairment," *Neurobiology of Aging*, vol. 27, no. 2, pp. 252–261, 2006.
- [56] T. Kamiya, Y. Katayama, F. Kashiwagi, and A. Terashi, "The role of bradykinin in mediating ischemic brain edema in rats," *Stroke*, vol. 24, no. 4, pp. 571–576, 1993.
- [57] A. Verkhratsky, R. K. Orkand, and H. Kettenmann, "Glial calcium: homeostasis and signaling function," *Physiological Reviews*, vol. 78, no. 1, pp. 99–141, 1998.

- [58] D. Regoli, N.-E. Rhaleb, S. Dion, and G. Drapeau, "New selective bradykinin receptor antagonists and bradykinin B2 receptor characterization," *Trends in Pharmacological Sciences*, vol. 11, no. 4, pp. 156–161, 1990.
- [59] K. D. Bhoola, C. D. Figueroa, and K. Worthly, "Bioregulation of kinins: kallikreins, kininogens, and kininases," *Pharmacological Reviews*, vol. 44, no. 1, pp. 1–80, 1992.
- [60] C.-W. Lin, S.-C. Shen, C.-C. Chien, L.-Y. Yang, L.-T. Shia, and Y.-C. Chen, "12-O-tetradecanoylphorbol-13-acetate-induced invasion/migration of glioblastoma cells through activating PKC α /ERK/NF- κ B-dependent MMP-9 expression," *Journal of Cellular Physiology*, vol. 225, no. 2, pp. 472–481, 2010.
- [61] A. Y. Abramov, J. Jacobson, F. Wientjes, J. Hotherhall, L. Canevari, and M. R. Duchon, "Expression and modulation of an NADPH oxidase in mammalian astrocytes," *The Journal of Neuroscience*, vol. 25, no. 40, pp. 9176–9184, 2005.
- [62] H.-L. Hsieh, H.-H. Wang, C.-Y. Wu, and C.-M. Yang, "Reactive oxygen species-dependent c-fos/activator protein 1 induction upregulates heme oxygenase-1 expression by bradykinin in brain astrocytes," *Antioxidants and Redox Signaling*, vol. 13, no. 12, pp. 1829–1844, 2010.
- [63] E. R. Levin, "Endothelins," *The New England Journal of Medicine*, vol. 333, no. 6, pp. 356–363, 1995.
- [64] S. Schinelli, "Pharmacology and physiopathology of the brain endothelin system: an overview," *Current Medicinal Chemistry*, vol. 13, no. 6, pp. 627–638, 2006.
- [65] F. Böhm and J. Pernow, "The importance of endothelin-1 for vascular dysfunction in cardiovascular disease," *Cardiovascular Research*, vol. 76, no. 1, pp. 8–18, 2007.
- [66] M. Hasselblatt, P. Lewczuk, B.-M. Löffler et al., "Role of the astrocytic ETB receptor in the regulation of extracellular endothelin-1 during hypoxia," *Glia*, vol. 34, no. 1, pp. 18–26, 2001.
- [67] S. D. Rogers, C. M. Peters, J. D. Pomonis, H. Hagiwara, J. R. Ghilardi, and P. W. Mantyh, "Endothelin B receptors are expressed by astrocytes and regulate astrocyte hypertrophy in the normal and injured CNS," *Glia*, vol. 41, no. 2, pp. 180–190, 2003.
- [68] A. C. Y. Lo, A. Y. S. Chen, V. K. L. Hung et al., "Endothelin-1 overexpression leads to further water accumulation and brain edema after middle cerebral artery occlusion via aquaporin 4 expression in astrocytic end-feet," *Journal of Cerebral Blood Flow and Metabolism*, vol. 25, no. 8, pp. 998–1011, 2005.
- [69] N. Didier, I. A. Romero, C. Créminon, A. Wijkhuisen, J. Grassi, and A. Mabondzo, "Secretion of interleukin-1 β by astrocytes mediates endothelin-1 and tumour necrosis factor- α effects on human brain microvascular endothelial cell permeability," *Journal of Neurochemistry*, vol. 86, no. 1, pp. 246–254, 2003.
- [70] S. J. Lee and S. Lee, "Toll-like receptors and inflammation in the CNS," *Current Drug Targets Inflammation & Allergy*, vol. 1, no. 2, pp. 181–191, 2002.
- [71] S.-Y. Kim, J.-G. Lee, W.-S. Cho et al., "Role of NADPH oxidase-2 in lipopolysaccharide-induced matrix metalloproteinase expression and cell migration," *Immunology and Cell Biology*, vol. 88, no. 2, pp. 197–204, 2010.
- [72] G. W. Konat, T. Kielian, and I. Marriott, "The role of Toll-like receptors in CNS response to microbial challenge," *Journal of Neurochemistry*, vol. 99, no. 1, pp. 1–12, 2006.
- [73] I. C. Sutcliffe and N. Shaw, "Atypical lipoteichoic acids of gram-positive bacteria," *Journal of Bacteriology*, vol. 173, no. 22, pp. 7065–7069, 1991.
- [74] X. Sáez-Llorens and G. H. McCracken Jr., "Bacterial meningitis in children," *The Lancet*, vol. 361, no. 9375, pp. 2139–2148, 2003.
- [75] S. C. Mullaly and P. Kubers, "The role of TLR2 in vivo following challenge with *Staphylococcus aureus* and prototypic ligands," *The Journal of Immunology*, vol. 177, no. 11, pp. 8154–8163, 2006.
- [76] J. A. Mitchell, M. J. Paul-Clark, G. W. Clarke, S. K. McMaster, and N. Cartwright, "Critical role of toll-like receptors and nucleotide oligomerisation domain in the regulation of health and disease," *Journal of Endocrinology*, vol. 193, no. 3, pp. 323–330, 2007.
- [77] A. Kinsner, V. Pilotto, S. Deininger et al., "Inflammatory neurodegeneration induced by lipoteichoic acid from *Staphylococcus aureus* is mediated by glia activation, nitrosative and oxidative stress, and caspase activation," *Journal of Neurochemistry*, vol. 95, no. 4, pp. 1132–1143, 2005.
- [78] S. Lehnardt, P. Henneke, E. Lien et al., "A mechanism for neurodegeneration induced by group B *Streptococci* through activation of the TLR2/MyD88 pathway in microglia," *The Journal of Immunology*, vol. 177, no. 1, pp. 583–592, 2006.
- [79] P. A. Carpentier, D. S. Duncan, and S. D. Miller, "Glial toll-like receptor signaling in central nervous system infection and autoimmunity," *Brain, Behavior, and Immunity*, vol. 22, no. 2, pp. 140–147, 2008.
- [80] M. Bsibsi, J. J. Bajramovic, E. van Duijvenvoorden et al., "Identification of soluble CD14 as an endogenous agonist for toll-like receptor 2 on human astrocytes by genome-scale functional screening of glial cell derived proteins," *Glia*, vol. 55, no. 5, pp. 473–482, 2007.
- [81] C. S. Jack, N. Arbour, J. Manusow et al., "TLR signaling tailors innate immune responses in human microglia and astrocytes," *The Journal of Immunology*, vol. 175, no. 7, pp. 4320–4330, 2005.
- [82] J. J. Neher and G. C. Brown, "Neurodegeneration in models of Gram-positive bacterial infections of the central nervous system," *Biochemical Society Transactions*, vol. 35, no. 5, pp. 1166–1167, 2007.
- [83] P. K. Chatterjee, K. Zacharowski, S. Cuzzocrea et al., "Lipoteichoic acid from *Staphylococcus aureus* reduces renal ischemia/reperfusion injury," *Kidney International*, vol. 62, no. 4, pp. 1249–1263, 2002.
- [84] G. Palacios and M. S. Oberste, "Enteroviruses as agents of emerging infectious diseases," *Journal of Neurovirology*, vol. 11, no. 5, pp. 424–433, 2005.
- [85] P. C. McMinn, "An overview of the evolution of enterovirus 71 and its clinical and public health significance," *FEMS Microbiology Reviews*, vol. 26, no. 1, pp. 91–107, 2002.
- [86] C.-C. Huang, C.-C. Liu, Y.-C. Chang, C.-Y. Chen, S.-T. Wang, and T.-F. Yeh, "Neurologic complications in children with enterovirus 71 infection," *The New England Journal of Medicine*, vol. 341, no. 13, pp. 936–942, 1999.
- [87] M. Ho, E.-R. Chen, K.-H. Hsu et al., "An epidemic of enterovirus 71 infection in Taiwan," *The New England Journal of Medicine*, vol. 341, no. 13, pp. 929–935, 1999.
- [88] W.-H. Tung, H.-L. Hsieh, I.-T. Lee, and C.-M. Yang, "Enterovirus 71 induces integrin β 1/EGFR-Rac1-dependent oxidative stress in SK-N-SH cells: role of HO-1/CO in viral replication," *Journal of Cellular Physiology*, vol. 226, no. 12, pp. 3316–3329, 2011.
- [89] U. K. Misra and J. Kalita, "Overview: Japanese encephalitis," *Progress in Neurobiology*, vol. 91, no. 2, pp. 108–120, 2010.

- [90] S.-L. Raung, S.-Y. Chen, S.-L. Liao, J.-H. Chen, and C.-J. Chen, "Tyrosine kinase inhibitors attenuate Japanese encephalitis virus-induced neurotoxicity," *Biochemical and Biophysical Research Communications*, vol. 327, no. 2, pp. 399–406, 2005.
- [91] M. K. Mishra, P. Koli, S. Bhowmick, and A. Basu, "Neuroprotection conferred by astrocytes is insufficient to protect animals from succumbing to Japanese encephalitis," *Neurochemistry International*, vol. 50, no. 5, pp. 764–773, 2007.
- [92] T. J. Montine, K. S. Montine, and L. L. Swift, "Central nervous system lipoproteins in Alzheimer's disease," *American Journal of Pathology*, vol. 151, no. 6, pp. 1571–1575, 1997.
- [93] J. N. Keller, K. B. Hanni, and W. R. Markesbery, "Oxidized low-density lipoprotein induces neuronal death: implications for calcium, reactive oxygen species, and caspases," *Journal of Neurochemistry*, vol. 72, no. 6, pp. 2601–2609, 1999.
- [94] F.-S. Shie, M. D. Neely, I. Maezawa et al., "Oxidized low-density lipoprotein is present in astrocytes surrounding cerebral infarcts and stimulates astrocyte interleukin-6 secretion," *American Journal of Pathology*, vol. 164, no. 4, pp. 1173–1181, 2004.
- [95] M. Uno, M. Harada, O. Takimoto et al., "Elevation of plasma oxidized LDL in acute stroke patients is associated with ischemic lesions depicted by DWI and predictive of infarct enlargement," *Neurological Research*, vol. 27, no. 1, pp. 94–102, 2005.
- [96] H.-H. Wang, H.-L. Hsieh, C.-Y. Wu, and C.-M. Yang, "Oxidized low-density lipoprotein-induced matrix metalloproteinase-9 expression via PKC- δ /p42/p44 MAPK/Elk-1 cascade in brain astrocytes," *Neurotoxicity Research*, vol. 17, no. 1, pp. 50–65, 2010.
- [97] P. Ten Dijke and C. S. Hill, "New insights into TGF- β -Smad signalling," *Trends in Biochemical Sciences*, vol. 29, no. 5, pp. 265–273, 2004.
- [98] J. Massagué, "How cells read TGF- β signals," *Nature Reviews Molecular Cell Biology*, vol. 1, no. 3, pp. 169–178, 2000.
- [99] S.-K. Leivonen, A. Chantry, L. Häkkinen, J. Han, and V.-M. Kähäri, "Smad3 mediates transforming growth factor- β -induced collagenase-3 (matrix metalloproteinase-13) expression in human gingival fibroblasts: evidence for cross-talk between Smad3 and p38 signaling pathways," *The Journal of Biological Chemistry*, vol. 277, no. 48, pp. 46338–46346, 2002.
- [100] K. Koli, M. Myllärniemi, J. Keski-Oja, and V. L. Kinnula, "Transforming growth factor- β activation in the lung: focus on fibrosis and reactive oxygen species," *Antioxidants and Redox Signaling*, vol. 10, no. 2, pp. 333–342, 2008.
- [101] F. Zipp and O. Aktas, "The brain as a target of inflammation: common pathways link inflammatory and neurodegenerative diseases," *Trends in Neurosciences*, vol. 29, no. 9, pp. 518–527, 2006.
- [102] A. Vezzani and T. Granata, "Brain inflammation in epilepsy: experimental and clinical evidence," *Epilepsia*, vol. 46, no. 11, pp. 1724–1743, 2005.
- [103] A. Simi, N. Tsakiri, P. Wang, and N. J. Rothwell, "Interleukin-1 and inflammatory neurodegeneration," *Biochemical Society Transactions*, vol. 35, no. 5, pp. 1122–1126, 2007.
- [104] V. W. Yong, C. A. Krekoski, P. A. Forsyth, R. Bell, and D. R. Edwards, "Matrix metalloproteinases and diseases of the CNS," *Trends in Neurosciences*, vol. 21, no. 2, pp. 75–80, 1998.
- [105] V. W. Yong, C. Power, P. Forsyth, and D. R. Edwards, "Metalloproteinases in biology and pathology of the nervous system," *Nature Reviews Neuroscience*, vol. 2, no. 7, pp. 502–511, 2001.
- [106] P. E. Gottschall and X. Yu, "Cytokines regulate gelatinase A and B (matrix metalloproteinase 2 and 9) activity in cultured rat astrocytes," *Journal of Neurochemistry*, vol. 64, no. 4, pp. 1513–1520, 1995.
- [107] W. J. Lee, C. Y. Shin, B. K. Yoo et al., "Induction of matrix metalloproteinase-9 (MMP-9) in lipopolysaccharide-stimulated primary astrocytes is mediated by extracellular signal-regulated protein kinase 1/2 (Erk1/2)," *Glia*, vol. 41, no. 1, pp. 15–24, 2003.
- [108] M. Hernández, M. L. Nieto, and M. Sánchez Crespo, "Cytosolic phospholipase A2 and the distinct transcriptional programs of astrocytoma cells," *Trends in Neurosciences*, vol. 23, no. 6, pp. 259–264, 2000.
- [109] I. Kudo and M. Murakami, "Phospholipase A₂ enzymes," *Prostaglandins and Other Lipid Mediators*, vol. 68–69, pp. 3–58, 2002.
- [110] J. Y. Park, M. H. Pillinger, and S. B. Abramson, "Prostaglandin E2 synthesis and secretion: the role of PGE2 synthases," *Clinical Immunology*, vol. 119, no. 3, pp. 229–240, 2006.
- [111] J. Xu, M. Chalimoniuk, Y. Shu et al., "Prostaglandin E2 production in astrocytes: regulation by cytokines, extracellular ATP, and oxidative agents," *Prostaglandins Leukotrienes and Essential Fatty Acids*, vol. 69, no. 6, pp. 437–448, 2003.
- [112] C. C. Leslie, "Properties and regulation of cytosolic phospholipase A₂," *The Journal of Biological Chemistry*, vol. 272, no. 27, pp. 16709–16712, 1997.
- [113] G. Y. Sun, J. Xu, M. D. Jensen et al., "Phospholipase A2 in astrocytes: responses to oxidative stress, inflammation, and G protein-coupled receptor agonists," *Molecular Neurobiology*, vol. 31, no. 1–3, pp. 27–41, 2005.
- [114] D. Stephenson, K. Rash, B. Smalstig et al., "Cytosolic phospholipase A2 is induced in reactive glia following different forms of neurodegeneration," *Glia*, vol. 27, no. 2, pp. 110–128, 1999.
- [115] M. T. Gentile, M. G. Reccia, P. P. Sorrentino et al., "Role of cytosolic calcium-dependent phospholipase A2 in Alzheimer's disease pathogenesis," *Molecular Neurobiology*, vol. 45, no. 3, pp. 596–604, 2012.
- [116] I. Szaingurten-Solodkin, N. Hadad, and R. Levy, "Regulatory role of cytosolic phospholipase A2 α in NADPH oxidase activity and in inducible nitric oxide synthase induction by aggregated A β 1–42 in microglia," *Glia*, vol. 57, no. 16, pp. 1727–1740, 2009.
- [117] C. Guo, J. Li, L. Myatt, X. Zhu, and K. Sun, "Induction of Gas contributes to the paradoxical stimulation of cytosolic phospholipase A2 α expression by cortisol in human amnion fibroblasts," *Molecular Endocrinology*, vol. 24, no. 5, pp. 1052–1061, 2010.
- [118] N. G. Bazan, V. Colangelo, and W. J. Lukiw, "Prostaglandins and other lipid mediators in Alzheimer's disease," *Prostaglandins and Other Lipid Mediators*, vol. 68–69, pp. 197–210, 2002.
- [119] C. S. Williams, M. Mann, and R. N. DuBois, "The role of cyclooxygenases in inflammation, cancer, and development," *Oncogene*, vol. 18, no. 55, pp. 7908–7916, 1999.
- [120] T. A. Samad, K. A. Moore, A. Sapirstein et al., "Interleukin-1 β -mediated induction of Cox-2 in the CNS contributes to inflammatory pain hypersensitivity," *Nature*, vol. 410, no. 6827, pp. 471–475, 2001.
- [121] T. Tanabe and N. Tohnai, "Cyclooxygenase isozymes and their gene structures and expression," *Prostaglandins and Other Lipid Mediators*, vol. 68–69, pp. 95–114, 2002.
- [122] M. Korotkova, M. Westman, K. R. Gheorghe et al., "Effects of antirheumatic treatments on the prostaglandin E2 biosynthetic pathway," *Arthritis and Rheumatism*, vol. 52, no. 11, pp. 3439–3447, 2005.

- [123] K. Ejima, M. D. Layne, I. M. Carvajal et al., "Cyclooxygenase-2-deficient mice are resistant to endotoxin-induced inflammation and death," *The FASEB Journal*, vol. 17, no. 10, pp. 1325–1327, 2003.
- [124] G. Tocco, J. Freire-Moar, S. S. Schreiber, S. H. Sakhi, P. S. Aisen, and G. M. Pasinetti, "Maturational regulation and regional induction of cyclooxygenase-2 in rat brain: implications for Alzheimer's disease," *Experimental Neurology*, vol. 144, no. 2, pp. 339–349, 1997.
- [125] W.-H. Tung, I.-T. Lee, H.-L. Hsieh, and C.-M. Yang, "EV71 induces COX-2 expression via c-Src/PDGFR/PI3K/Akt/p42/p44 MAPK/AP-1 and NF- κ B in rat brain astrocytes," *Journal of Cellular Physiology*, vol. 224, no. 2, pp. 376–386, 2010.
- [126] W.-H. Tung, H.-L. Hsieh, I.-T. Lee, and C.-M. Yang, "Enterovirus 71 modulates a COX-2/PGE2/cAMP-dependent viral replication in human neuroblastoma cells: role of the c-Src/EGFR/p42/p44 MAPK/CREB signaling pathway," *Journal of Cellular Biochemistry*, vol. 112, no. 2, pp. 559–570, 2011.
- [127] H. L. Hsieh, C. C. Lin, H. J. Chan, C. M. Yang, and C. M. Yang, "c-Src-dependent EGF receptor transactivation contributes to ET-1-induced COX-2 expression in brain microvascular endothelial cells," *Journal of Neuroinflammation*, vol. 9, p. 152, 2012.
- [128] D. S. Kim, J. Y. Kim, and Y. Han, "Curcuminoids in neurodegenerative diseases," *Recent Patents on CNS Drug Discovery*, vol. 7, no. 3, pp. 184–204, 2012.
- [129] K. J. Smith and H. Lassmann, "The role of nitric oxide in multiple sclerosis," *The Lancet Neurology*, vol. 1, no. 4, pp. 232–241, 2002.
- [130] L. H. Barbeito, M. Pehar, P. Cassina et al., "A role for astrocytes in motor neuron loss in amyotrophic lateral sclerosis," *Brain Research Reviews*, vol. 47, no. 1–3, pp. 263–274, 2004.
- [131] R. N. Saha and K. Pahan, "Regulation of inducible nitric oxide synthase gene in glial cells," *Antioxidants and Redox Signaling*, vol. 8, no. 5–6, pp. 929–947, 2006.
- [132] E. Galea, D. L. Feinstein, and D. J. Reis, "Induction of calcium-independent nitric oxide synthase activity in primary rat glial cultures," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 22, pp. 10945–10949, 1992.
- [133] S. Parmentier-Batteur, G. A. Bohme, D. Lerouet et al., "Anti-sense oligodeoxynucleotide to inducible nitric oxide synthase protects against transient focal cerebral ischemia-induced brain injury," *Journal of Cerebral Blood Flow and Metabolism*, vol. 21, no. 1, pp. 15–21, 2001.
- [134] J. Li, O. Baud, T. Vartanian, J. J. Volpe, and P. A. Rosenberg, "Peroxynitrite generated by inducible nitric oxide synthase and NADPH oxidase mediates microglial toxicity to oligodendrocytes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 28, pp. 9936–9941, 2005.
- [135] S.-H. Choi, E. H. Joe, S. U. Kim, and B. K. Jin, "Thrombin-induced microglial activation produces degeneration of nigral dopaminergic neurons in vivo," *The Journal of Neuroscience*, vol. 23, no. 13, pp. 5877–5886, 2003.
- [136] T. A. Springer, "Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm," *Cell*, vol. 76, no. 2, pp. 301–314, 1994.
- [137] J. M. Cook-Mills, "VCAM-1 signals during lymphocyte migration: role of reactive oxygen species," *Molecular Immunology*, vol. 39, no. 9, pp. 499–508, 2002.
- [138] L. Osborn, C. Hession, R. Tizard et al., "Direct expression cloning of vascular cell adhesion molecule 1, a cytokine-induced endothelial protein that binds to lymphocytes," *Cell*, vol. 59, no. 6, pp. 1203–1211, 1989.
- [139] M. Michalska, L. Machtoub, H. D. Manthey et al., "Visualization of vascular inflammation in the atherosclerotic mouse by ultrasmall superparamagnetic iron oxide vascular cell adhesion molecule-1-specific nanoparticles," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 32, no. 10, pp. 2350–2357, 2012.
- [140] C. Tang, H.-L. Xue, C.-L. Bai, and R. Fu, "Regulation of adhesion molecules expression in TNF- α -stimulated brain microvascular endothelial cells by tanshinone IIA: involvement of NF- κ B and ROS generation," *Phytotherapy Research*, vol. 25, no. 3, pp. 376–380, 2011.
- [141] H. Y. Song, J. Ryu, S. M. Ju et al., "Extracellular HIV-1 Tat enhances monocyte adhesion by up-regulation of ICAM-1 and VCAM-1 gene expression via ROS-dependent NF- κ B activation in astrocytes," *Experimental and Molecular Medicine*, vol. 39, no. 1, pp. 27–37, 2007.
- [142] M. R. Jacquier-Sarlin, K. Fuller, A. T. Dinh-Xuan, M.-J. Richard, and B. S. Polla, "Protective effects of hsp70 in inflammation," *Experientia*, vol. 50, no. 11–12, pp. 1031–1038, 1994.
- [143] P. Aridon, F. Geraci, G. Turturici, M. D'amelio, G. Savettieri, and G. Sconzo, "Protective role of heat shock proteins in Parkinson's disease," *Neurodegenerative Diseases*, vol. 8, no. 4, pp. 155–168, 2011.
- [144] W. Luo, W. Sun, T. Taldone, A. Rodina, and G. Chiosis, "Heat shock protein 90 in neurodegenerative diseases," *Molecular Neurodegeneration*, vol. 5, no. 1, article 24, 2010.
- [145] S. Patury, Y. Miyata, and J. E. Gestwicki, "Pharmacological targeting of the Hsp70 chaperone," *Current Topics in Medicinal Chemistry*, vol. 9, no. 15, pp. 1337–1351, 2009.
- [146] T. Yamashima, "Hsp70.1 and related lysosomal factors for necrotic neuronal death," *Journal of Neurochemistry*, vol. 120, no. 4, pp. 477–494, 2012.
- [147] J. M. Kyriakis and J. Avruch, "Mammalian mitogen-activated protein kinase signal transduction pathways activated by stress and inflammation," *Physiological Reviews*, vol. 81, no. 2, pp. 807–869, 2001.
- [148] E. A. Irving and M. Bamford, "Role of mitogen- and stress-activated kinases in ischemic injury," *Journal of Cerebral Blood Flow and Metabolism*, vol. 22, no. 6, pp. 631–647, 2002.
- [149] S. J. Harper and P. Lograsso, "Signalling for survival and death in neurones: the role of stress-activated kinases, JNK and p38," *Cellular Signalling*, vol. 13, no. 5, pp. 299–310, 2001.
- [150] K. Hensley, R. A. Floyd, N.-Y. Zheng et al., "p38 Kinase is activated in the Alzheimer's disease brain," *Journal of Neurochemistry*, vol. 72, no. 5, pp. 2053–2058, 1999.
- [151] J. Hu, K. T. Akama, G. A. Krafft, B. A. Chromy, and L. J. van Eldik, "Amyloid- β peptide activates cultured astrocytes: morphological alterations, cytokine induction and nitric oxide release," *Brain Research*, vol. 785, no. 2, pp. 195–206, 1998.
- [152] S. Deb, J. W. Zhang, and P. E. Gottschall, " β -amyloid induces the production of active, matrix-degrading proteases in cultured rat astrocytes," *Brain Research*, vol. 970, no. 1–2, pp. 205–213, 2003.
- [153] Y. Yarden and M. X. Sliwkowski, "Untangling the ErbB signalling network," *Nature Reviews Molecular Cell Biology*, vol. 2, no. 2, pp. 127–137, 2001.
- [154] H. Daub, F. U. Weiss, C. Wallasch, and A. Ullrich, "Role of transactivation of the EGF receptor in signalling by G-protein-coupled receptors," *Nature*, vol. 379, no. 6565, pp. 557–560, 1996.
- [155] N. Prenzel, E. Zwick, H. Daub et al., "EGF receptor transactivation by G-protein-coupled receptors requires metalloproteinase

- cleavage of proHB-EGF," *Nature*, vol. 402, no. 6764, pp. 884–888, 1999.
- [156] F. S. Lee and M. V. Chao, "Activation of Trk neurotrophin receptors in the absence of neurotrophins," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 98, no. 6, pp. 3555–3560, 2001.
- [157] A. Herrlich, H. Daub, A. Knebel et al., "Ligand-independent activation of platelet-derived growth factor receptor is a necessary intermediate in lysophosphatidic, acid-stimulated mitogenic activity in L cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 15, pp. 8985–8990, 1998.
- [158] F. G. Buchanan, D. Wang, F. Bargiacchi, and R. N. DuBois, "Prostaglandin E2 regulates cell migration via the intracellular activation of the epidermal growth factor receptor," *The Journal of Biological Chemistry*, vol. 278, no. 37, pp. 35451–35457, 2003.
- [159] T. Tanimoto, Z.-G. Jin, and B. C. Berk, "Transactivation of vascular endothelial growth factor (VEGF) receptor Flk-1/KDR is involved in sphingosine 1-phosphate-stimulated phosphorylation of Akt and endothelial nitric-oxide synthase (eNOS)," *The Journal of Biological Chemistry*, vol. 277, no. 45, pp. 42997–43001, 2002.
- [160] G. Neufeld, T. Cohen, S. Gengrinovitch, and Z. Poltorak, "Vascular endothelial growth factor (VEGF) and its receptors," *The FASEB Journal*, vol. 13, no. 1, pp. 9–22, 1999.
- [161] H. Lei and A. Kazlauskas, "Growth factors outside of the platelet-derived growth factor (PDGF) family employ reactive oxygen species/Src family kinases to activate PDGF receptor α and thereby promote proliferation and survival of cells," *The Journal of Biological Chemistry*, vol. 284, no. 10, pp. 6329–6336, 2009.
- [162] K. C.-W. Chen, Y. Zhou, K. Xing, K. Krysan, and M. F. Lou, "Platelet derived growth factor (PDGF)-induced reactive oxygen species in the lens epithelial cells: the redox signaling," *Experimental Eye Research*, vol. 78, no. 6, pp. 1057–1067, 2004.
- [163] B. Liu and A. H. Neufeld, "Activation of epidermal growth factor receptors in astrocytes: from development to neural injury," *Journal of Neuroscience Research*, vol. 85, no. 16, pp. 3523–3529, 2007.
- [164] A. Ptasznik, E. R. Prossnitz, D. Yoshikawa, A. Smrcka, A. E. Traynor-Kaplan, and G. M. Bokoch, "A tyrosine kinase signaling pathway accounts for the majority of phosphatidylinositol 3,4,5-trisphosphate formation in chemoattractant-stimulated human neutrophils," *The Journal of Biological Chemistry*, vol. 271, no. 41, pp. 25204–25207, 1996.
- [165] C. Angeloni, E. Motori, D. Fabbri et al., "H₂O₂ preconditioning modulates phase II enzymes through p38 MAPK and PI3K/Akt activation," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 300, no. 6, pp. H2196–H2205, 2011.
- [166] J. Pan, Q. Chang, X. Wang et al., "Reactive oxygen species-activated Akt/ASK1/p38 signaling pathway in nickel compound-induced apoptosis in BEAS 2B cells," *Chemical Research in Toxicology*, vol. 23, no. 3, pp. 568–577, 2010.
- [167] A. S. Jr Baldwin, "The NF- κ B and I κ B proteins: new discoveries and insights," *Annual Review of Immunology*, vol. 14, pp. 649–683, 1996.
- [168] J. J. Haddad, "Oxygen-sensitive pro-inflammatory cytokines, apoptosis signaling and redox-responsive transcription factors in development and pathophysiology," *Cytokines, Cellular and Molecular Therapy*, vol. 7, no. 1, pp. 1–14, 2002.
- [169] P. J. Barnes and M. Karin, "Nuclear factor- κ B—a pivotal transcription factor in chronic inflammatory diseases," *The New England Journal of Medicine*, vol. 336, no. 15, pp. 1066–1071, 1997.
- [170] W. Eberhardt, A. Huwiler, K.-F. Beck, S. Walpen, and J. Pfeilschifter, "Amplification of IL-1 β -induced matrix metalloproteinase-9 expression by superoxide in rat glomerular mesangial cells is mediated by increased activities of NF- κ B and activating protein-1 and involves activation of the mitogen-activated protein kinase pathways," *The Journal of Immunology*, vol. 165, no. 10, pp. 5788–5797, 2000.
- [171] S. Y. Kim, K.-A. Moon, H.-Y. Jo et al., "Anti-inflammatory effects of apocynin, an inhibitor of NADPH oxidase, in airway inflammation," *Immunology and Cell Biology*, vol. 90, no. 4, pp. 441–448, 2012.
- [172] S.-E. Cheng, C.-C. Lin, I.-T. Lee, C.-K. Hsu, Y. R. Kou, and C.-M. Yang, "Cigarette smoke extract regulates cytosolic phospholipase A2 expression via NADPH oxidase/MAPKs/AP-1 and p300 in human tracheal smooth muscle cells," *Journal of Cellular Biochemistry*, vol. 112, no. 2, pp. 589–599, 2011.
- [173] H. M. Chan and N. B. La Thangue, "p300/CBP proteins: HATs for transcriptional bridges and scaffolds," *Journal of Cell Science*, vol. 114, no. 13, pp. 2363–2373, 2001.
- [174] H. Asahara, S. Tartare-Deckert, T. Nakagawa et al., "Dual roles of p300 in chromatin assembly and transcriptional activation in cooperation with nucleosome assembly protein 1 in vitro," *Molecular and Cellular Biology*, vol. 22, no. 9, pp. 2974–2983, 2002.
- [175] H. Ma, C. Nguyen, K.-S. Lee, and M. Kahn, "Differential roles for the coactivators CBP and p300 on TCF/ β -catenin-mediated survivin gene expression," *Oncogene*, vol. 24, no. 22, pp. 3619–3631, 2005.